

# Serial Section Electron Microscopy

## From Heroic to Routine

Contributions From:

Kristen Harris

Josef Spacek

John Fiala

Special thanks to

Pasko Rakic

Bitao Shi

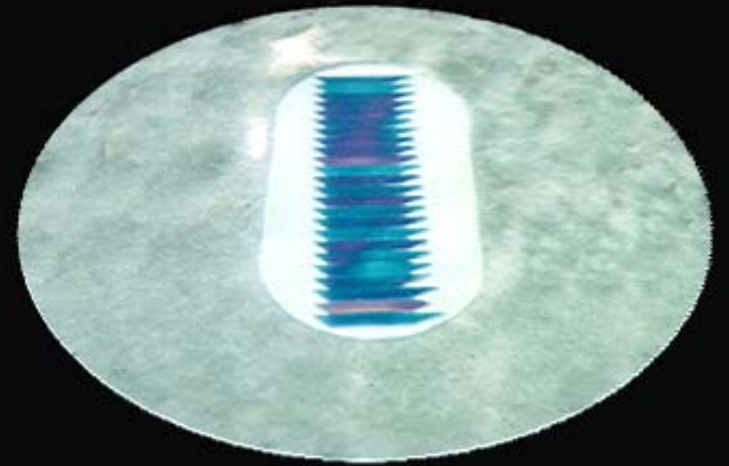
Elizabeth Perry

Robert Smith



**Josef Spacek**

# Some Heroes





# Aligned Outlines



# Solid Models

Schwann Cell Process



Synapse on Dendritic Spine





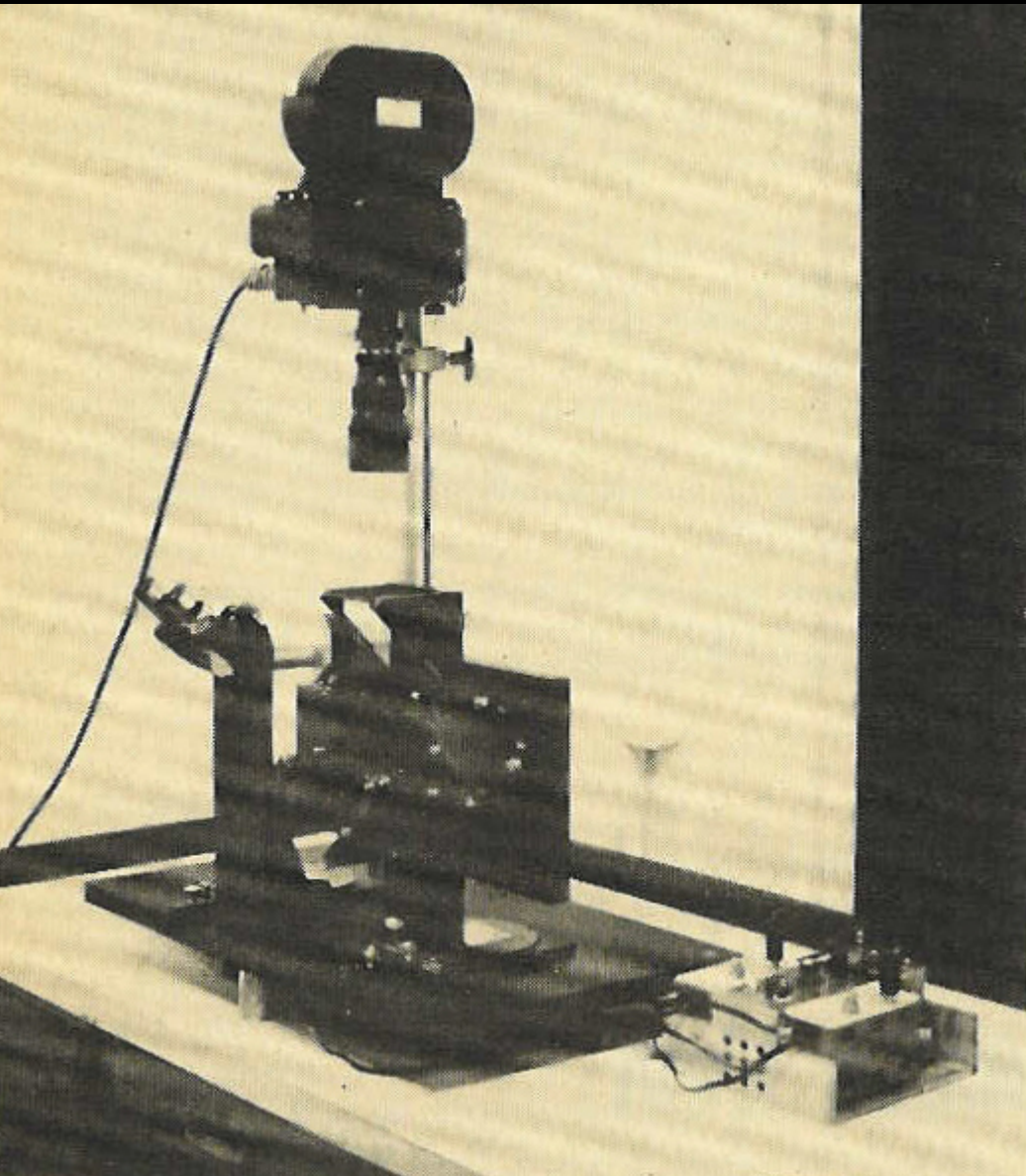


# Photomicrographs

## Laid End to End!

Kip Riley on the roof of  
Children's Hospital, Boston  
Mid 1970's – Radial Glia  
Reconstructions

Courtesy Pasko Rakic

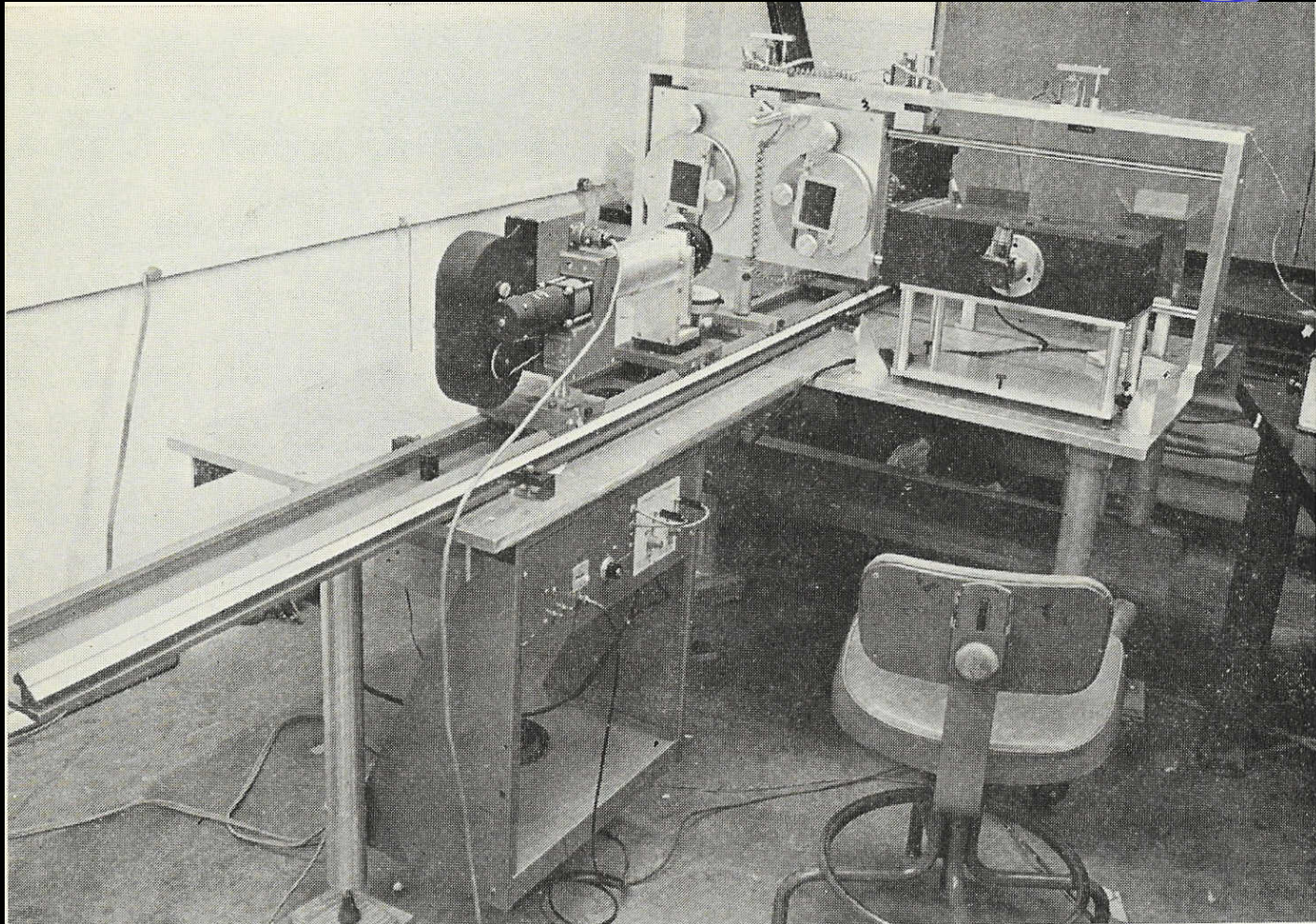


# **Cine Alignment of Serial Images Using Strobe Lights**

1972 Levinthal and  
Ware,— Nature 236:  
207-210)



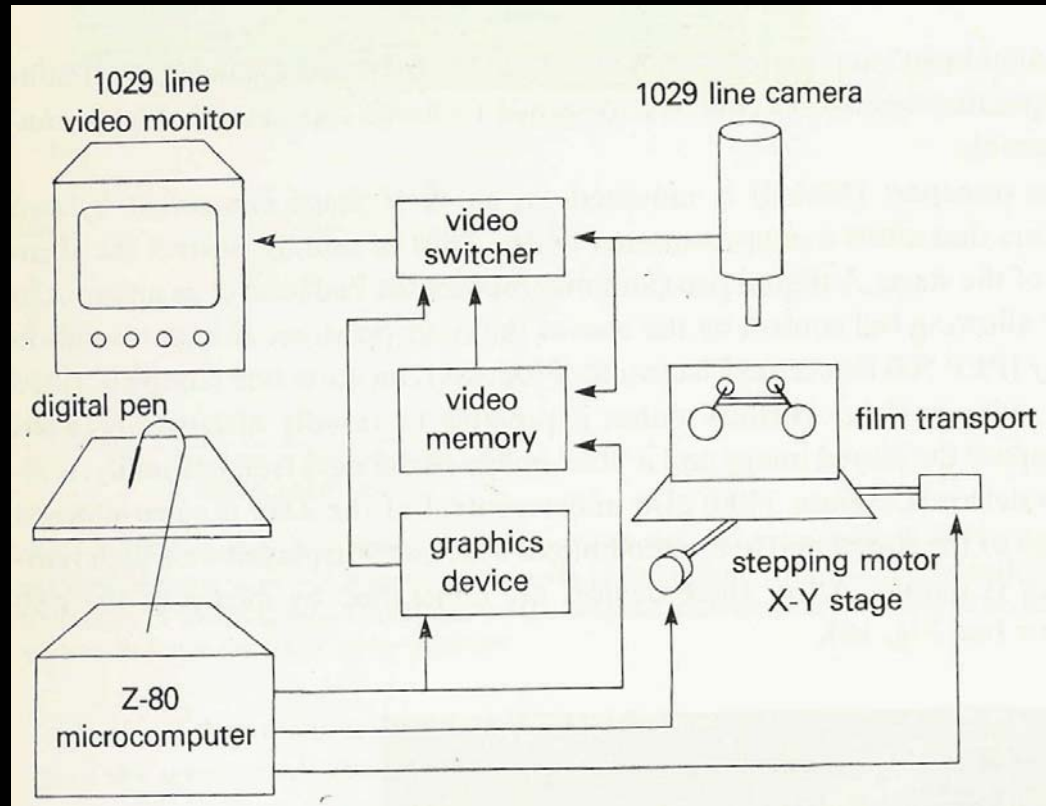
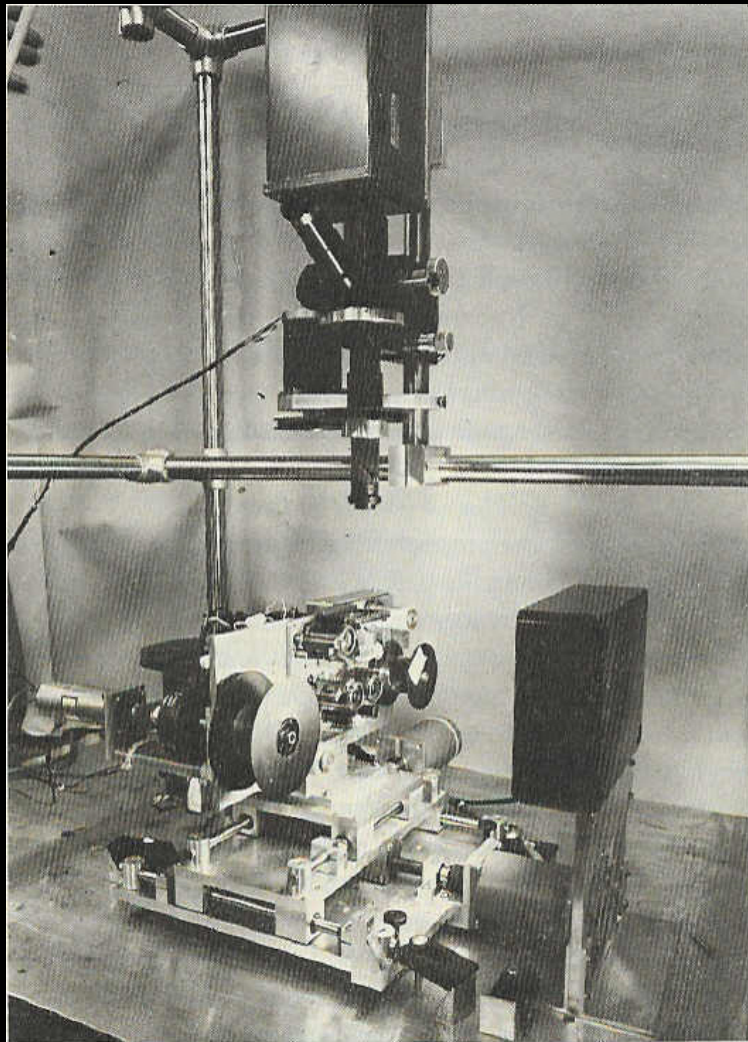
# Movie-making



1980 Stevens et al., Brain Res. Rev. 2:265



# Movie Transport and Computer-based Alignment

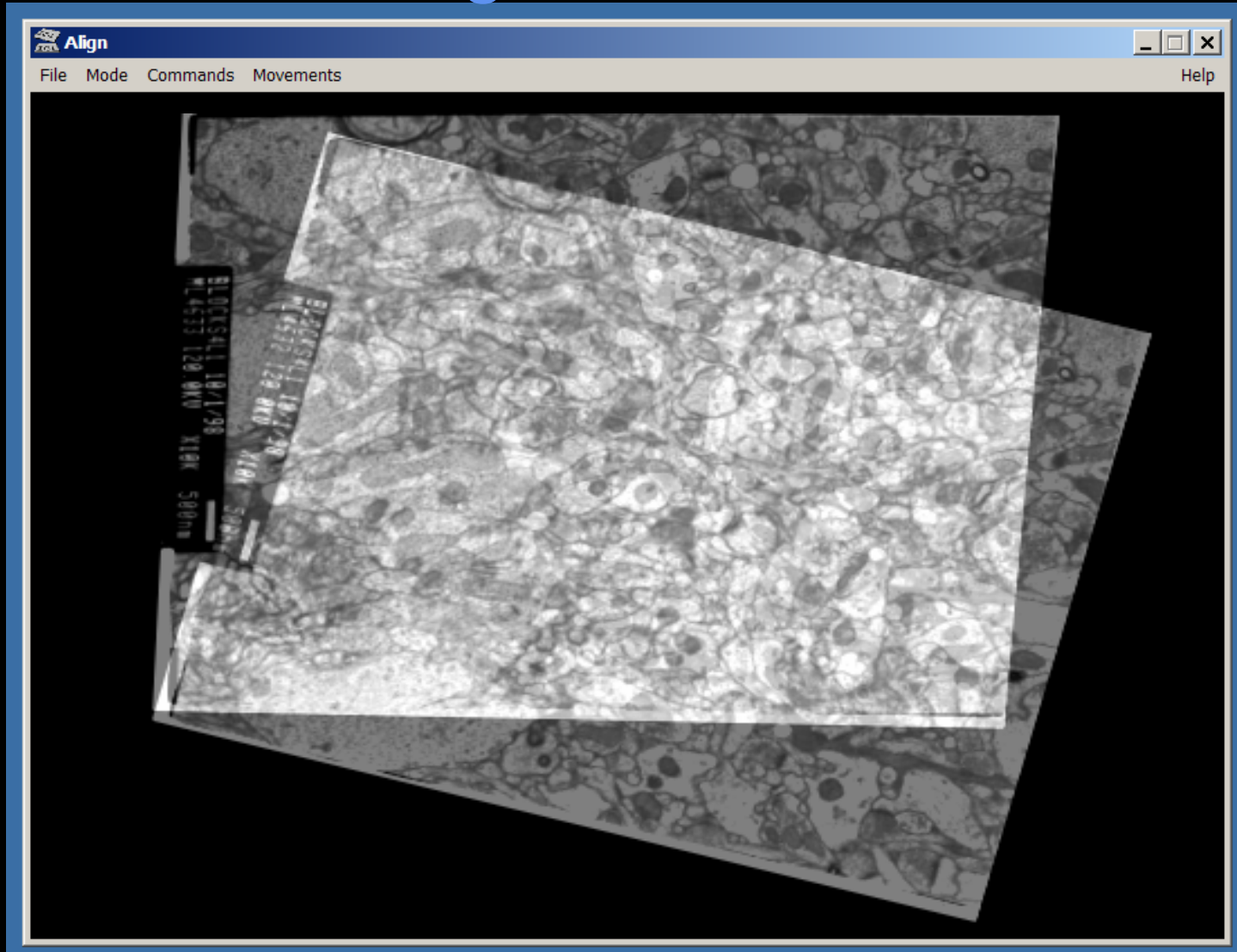


1980 - Stevens et al.,  
Brain Res. Rev. 2:265

# Sharing the Excitement



# *IGL Align* (1997) Alignment by Image Movement





# Routine Methods for Serial Section Electron Microscopy and 3D Reconstruction

Kristen M. Harris

Are POSTED AT:

SynapseWeb: [synapses.mcg.edu](http://synapses.mcg.edu)

Acknowledgements:

Libby Perry – “NEVER CUT CORNERS”!

Robert Smith

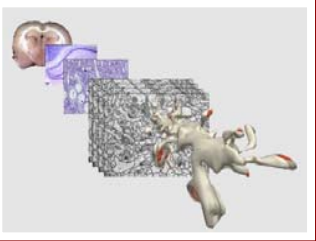
Marcia Feinberg

Jamie Hurlburt

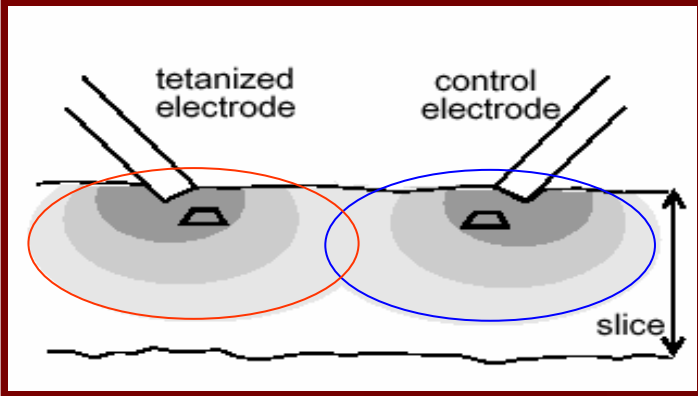
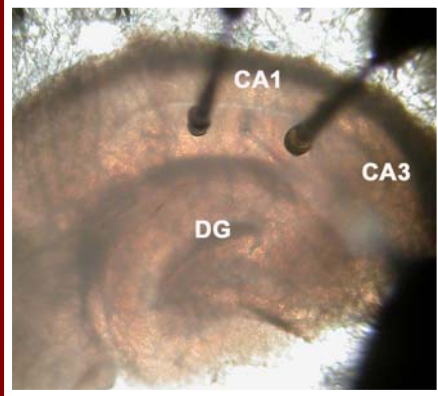
# Laboratory of Synapse Structure and Function

Synapses and Cognitive Neuroscience Center

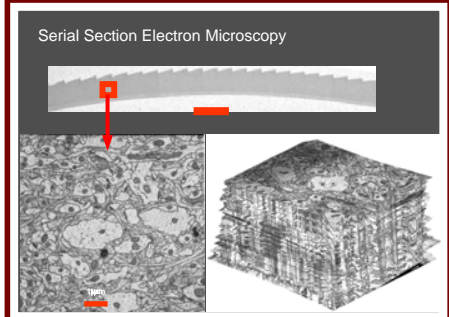
Kristen M. Harris, PhD, Principal Investigator



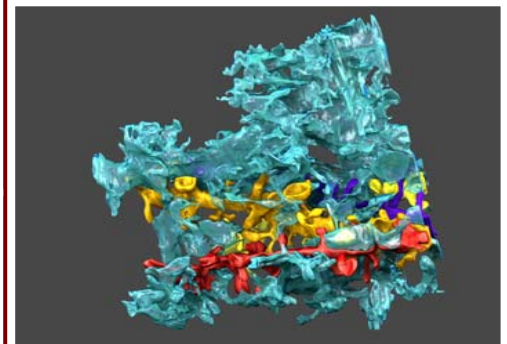
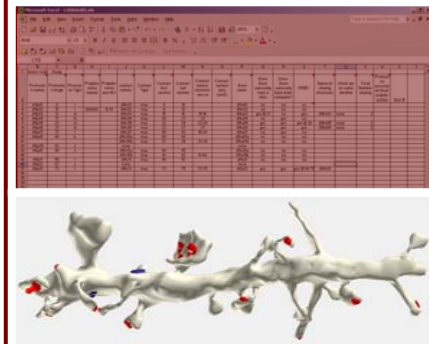
**Physiology and Processing**



**Sectioning and Microscopy**



**Tracing, Data Analysis & 3D Reconstruction**



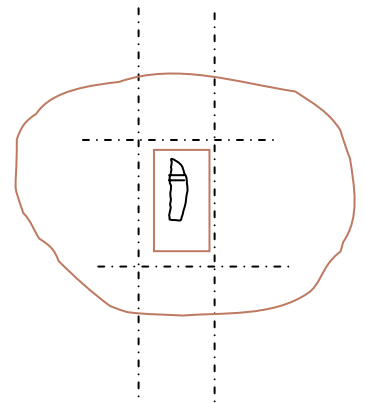
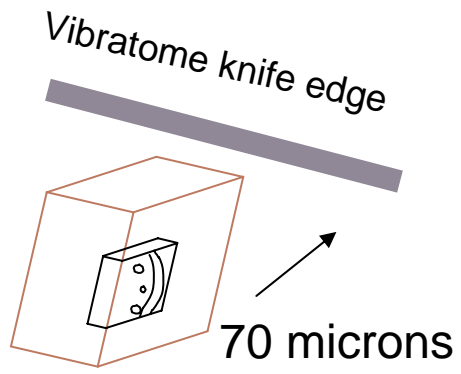
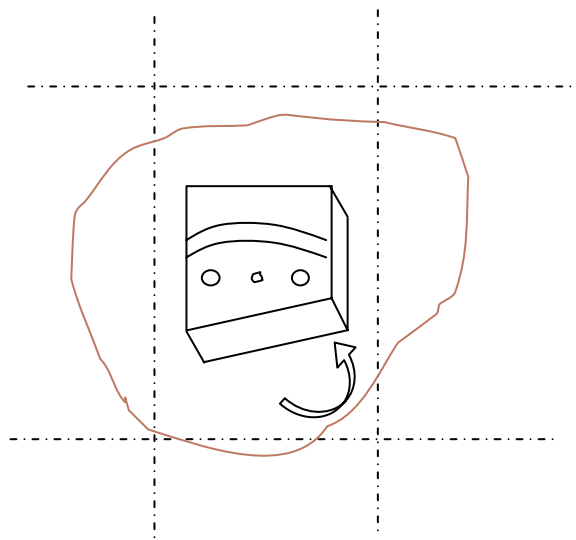
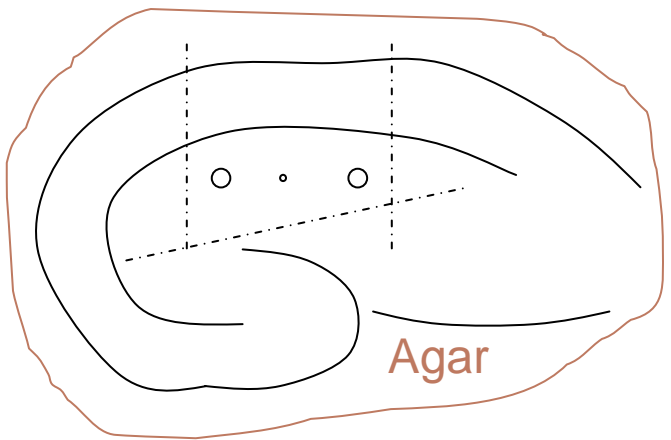
# At end of the experiment



- Remove electrodes
- Place slice on net, upside down onto a glass ring in fixative in a MW-oven
  - 6% glut, 2% para, in 0.1 M Cacodylate buffer with 2mM  $\text{Ca}^{2+}$  and 4mM  $\text{Mg}^{2+}$
- MW for about 8-20 seconds (700 Watts full power)
  - final temp  $\sim 37^{\circ}\text{C}$ , not  $> 45^{\circ}\text{C}$
- Fix overnight
- Vibratome and process next day!



# Trimming a region of interest



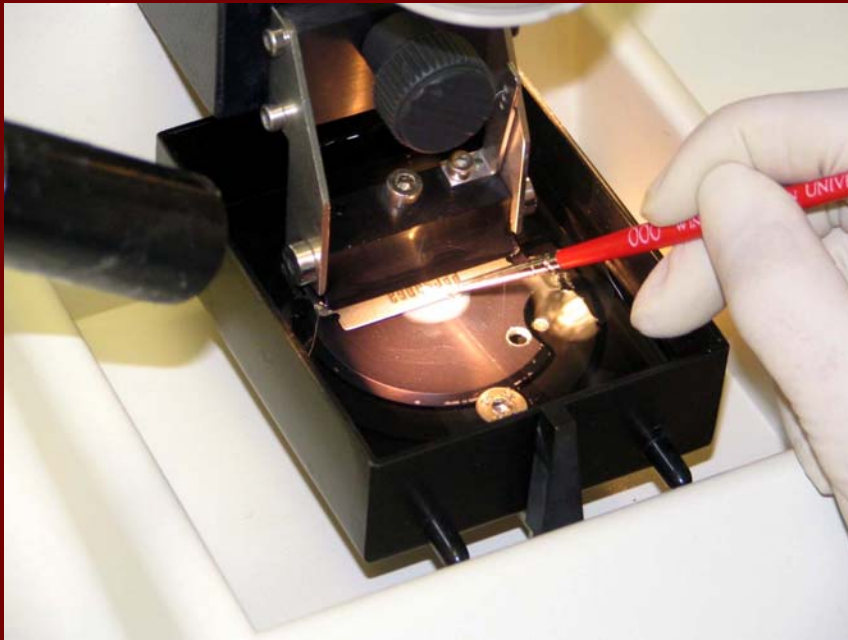
Re-embed to support and orient

# Vibratome at 70 microns



Ensures uniform penetration of osmium into the tissue

# Transfer 70 micron slices sequentially to 24 well TC dishes

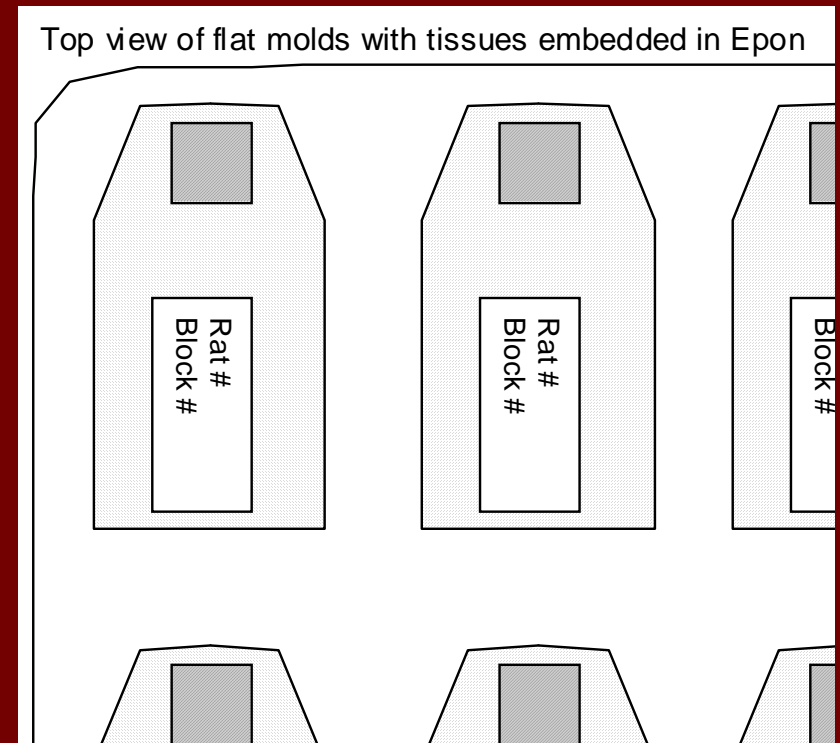


Collect sections with brush on corner of excess Agar

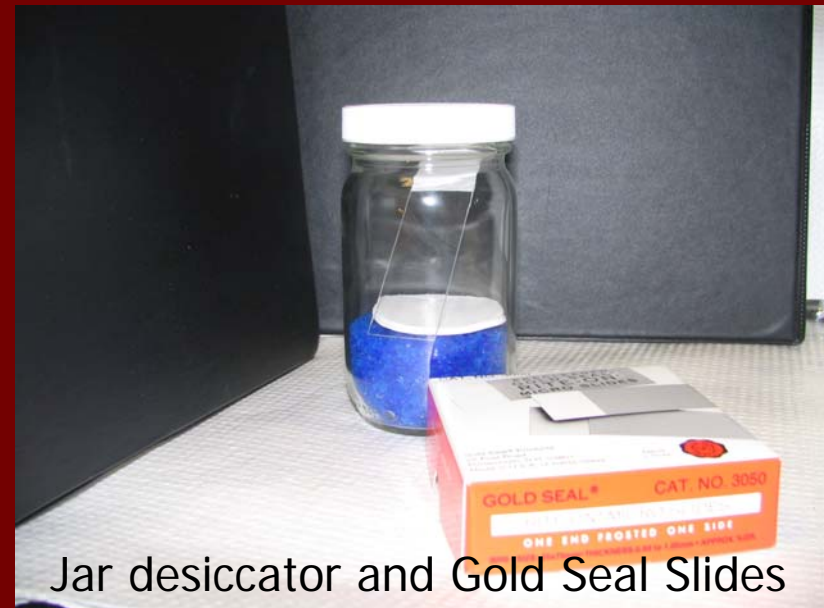
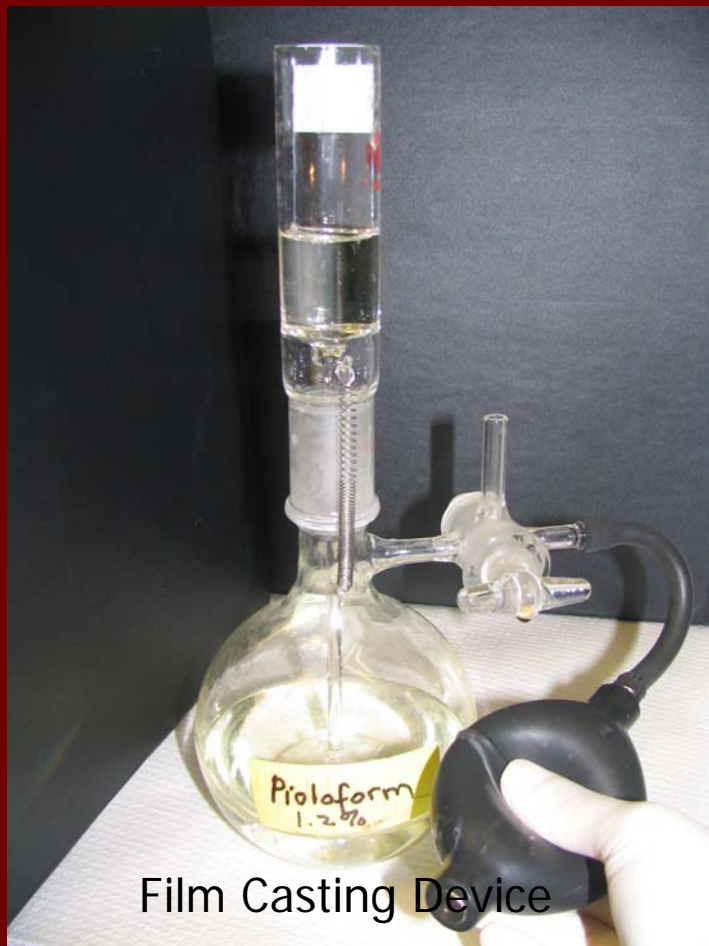


# Example Processing

- Osmium - K-Fe reduced then 1%
  - Optimizes membranes
- UA during dehydration in ethanols
  - Optimizes polyribosomes
- Acetone (or Propylene oxide?)
  - Acetone - translucent -measure
  - PO – black tissue
- Epoxy Resin (LX112) in Coffin Mold
- **Cure at 60 °C - 48 hrs!**
- Resurface with Epon/Aclar
  - To see through block in LM
- Details and “others” at:
  - [synapses.mcg.edu](http://synapses.mcg.edu) - lab protocols



# Make Pioloform-coated Slot Grids within 24 hours prior to serial sectioning



# People who make this happen



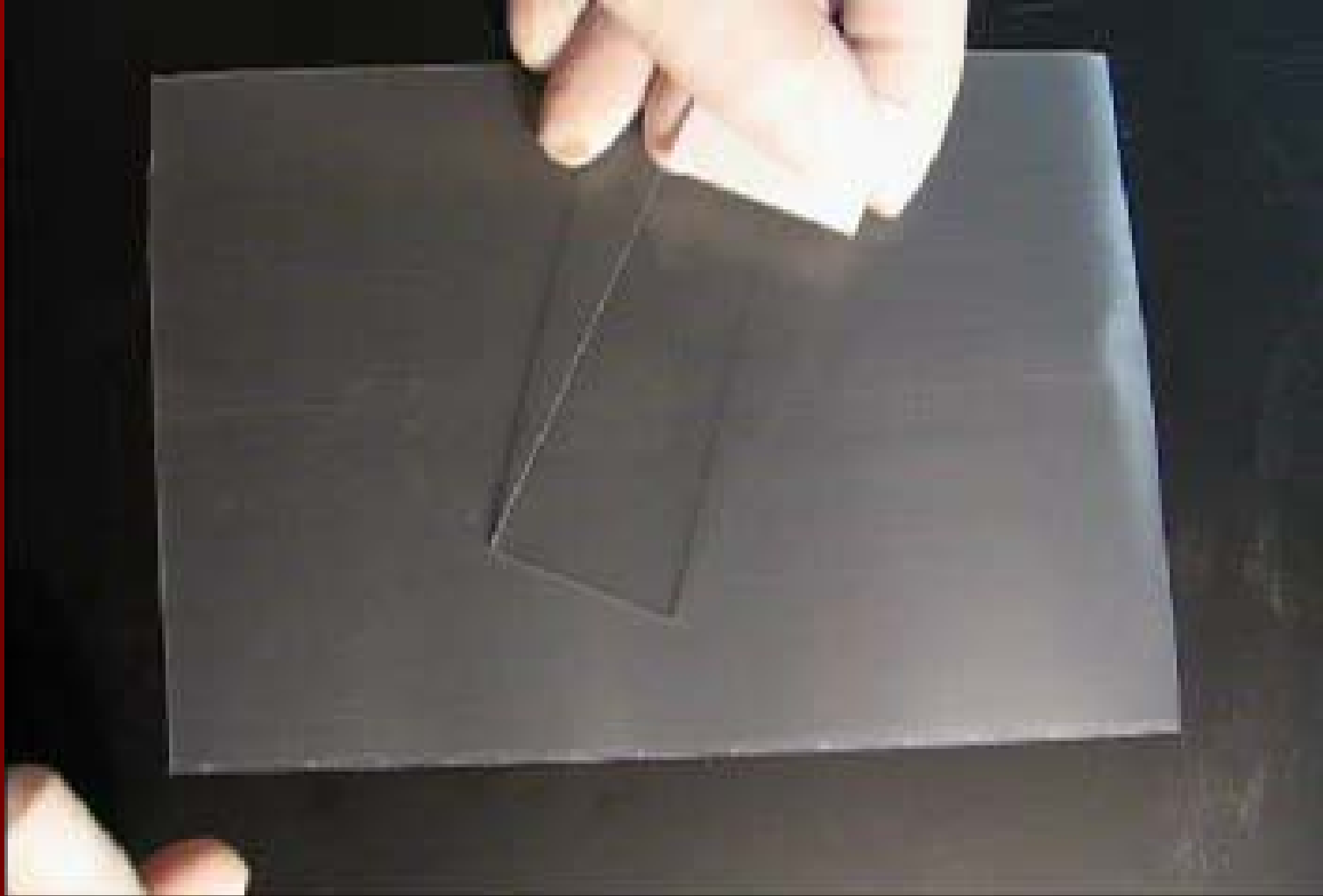
Robert Smith



Libby Perry

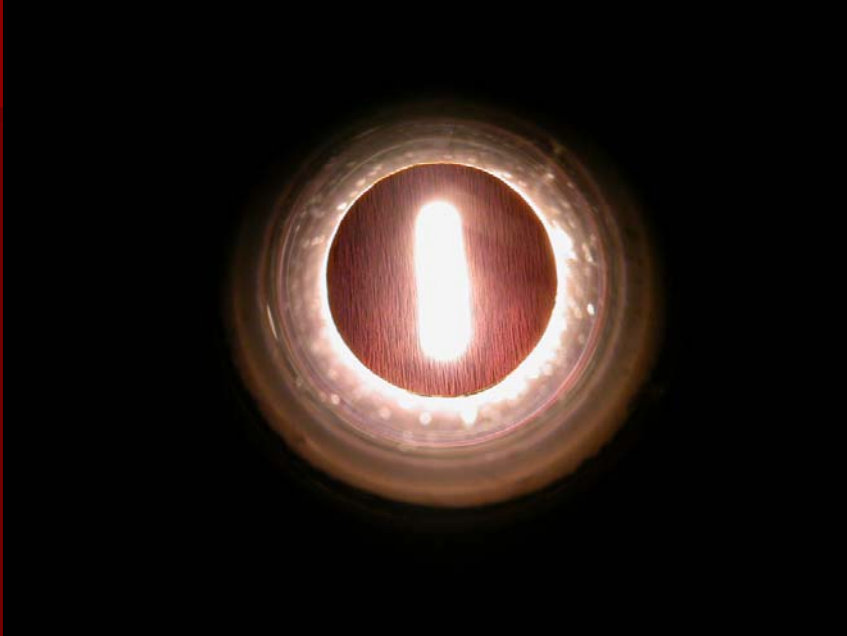


# Grid coating



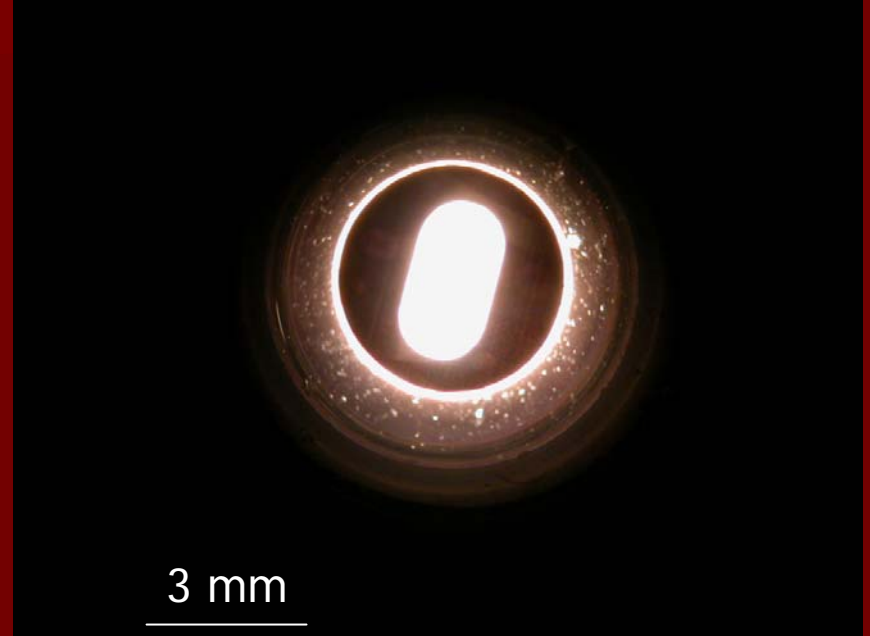
score coated slide and float off pioloform film

# Synaptek™ Slot Grids



**0.5 mm width**

**More stable, more difficult to  
center a long ribbon**



**1.0 mm width**

**Less stable, easier to  
center a long ribbon**

(stiff - beryllium-copper, 4 mil (100 $\mu$ m), from EMS or Ted Pella)

Place grids notch side up on silver colored film

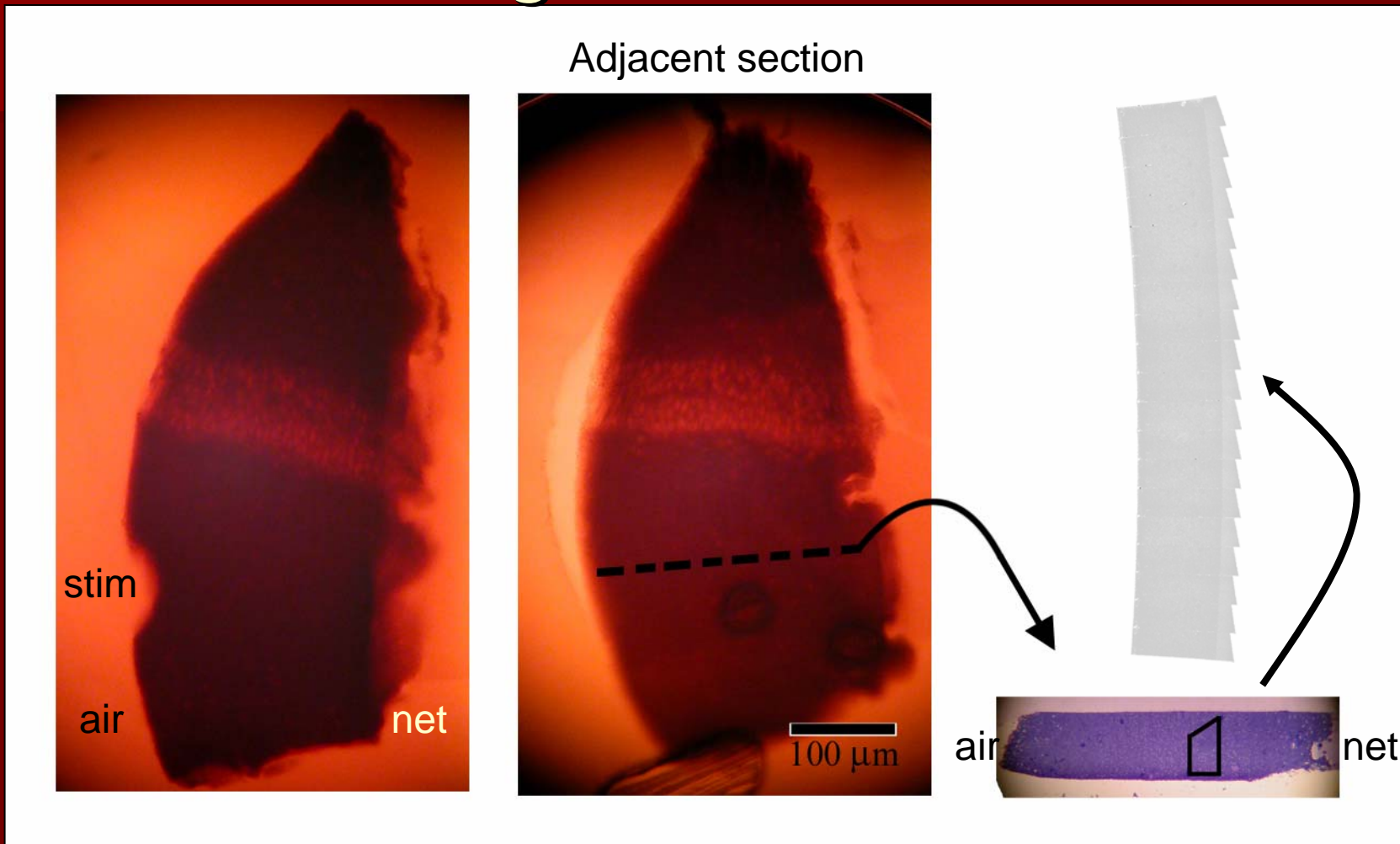




# Stretch and secure film on grids



# Goal: Uniformly thin serial sections in Region of Interest

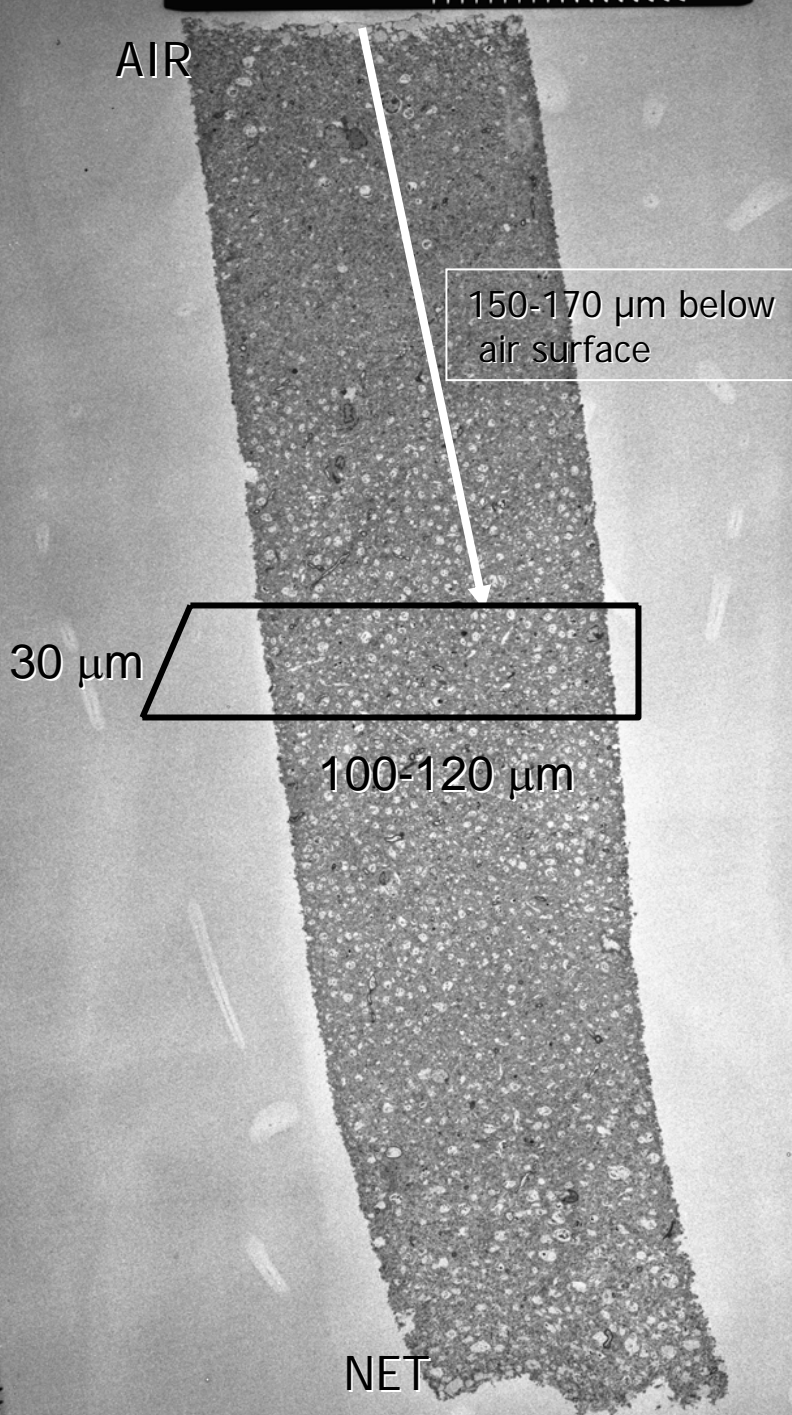


70 micron sections  
viewed through epoxy resin

Next Slides  
show steps

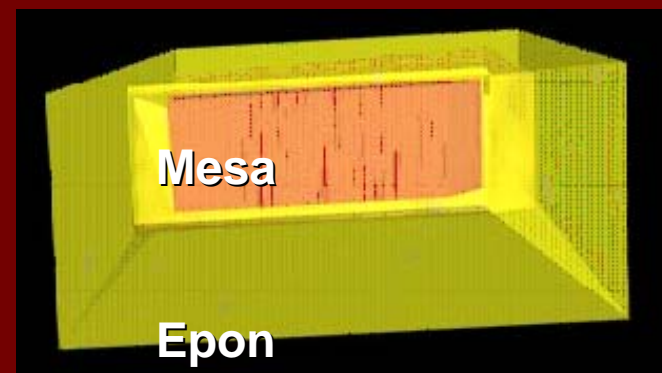
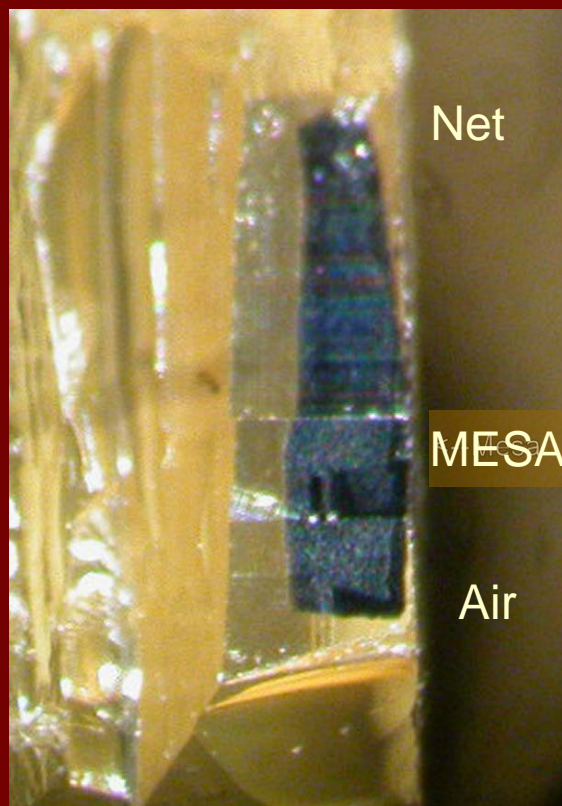
# Test thin across slice

- Cut and visualize test thin across depth of slice
- Evaluate tissue for good ultrastructure
- Identify where to place trapezoid for series
- Blind as to condition of course!



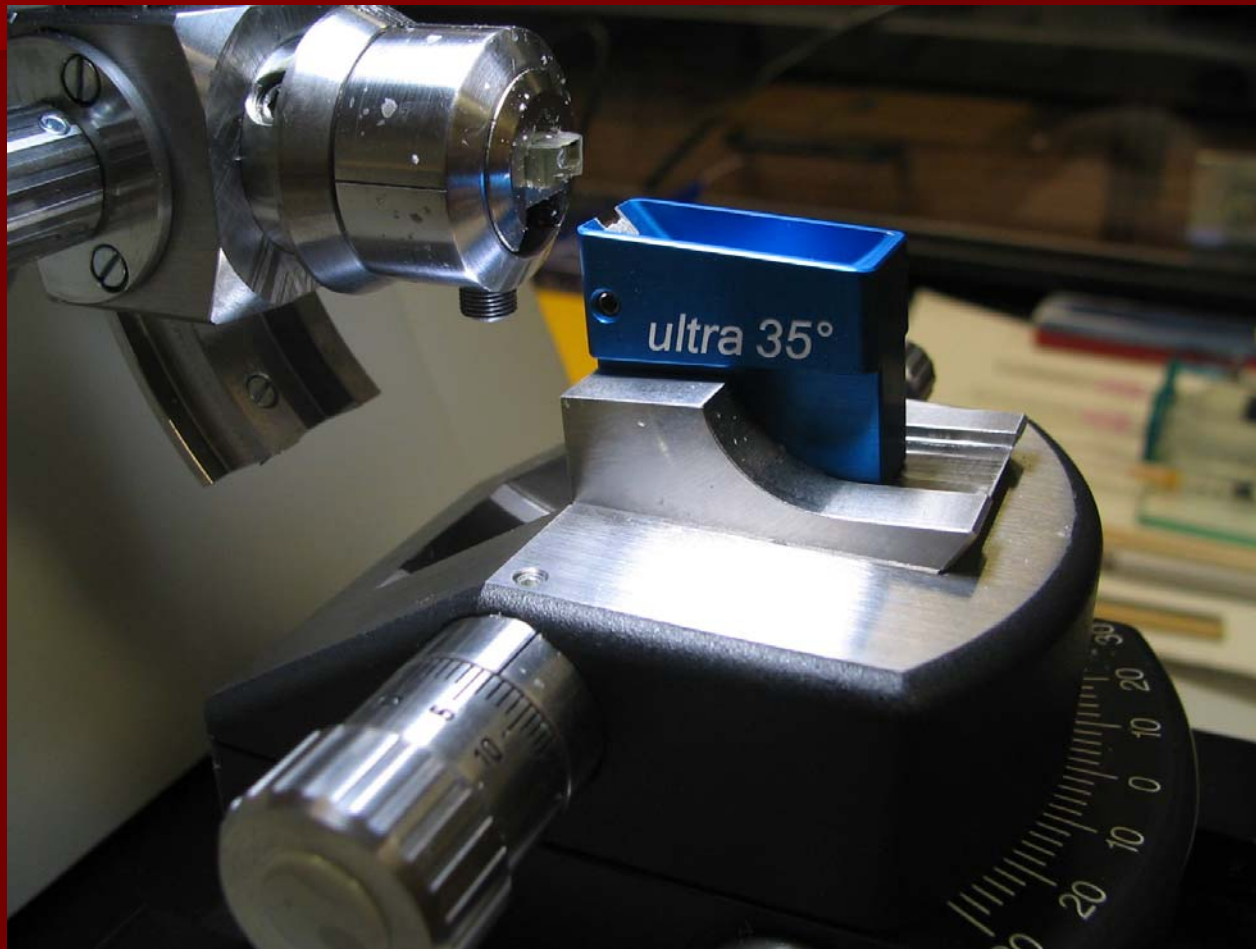


# Trim Mesa and Trapezoid

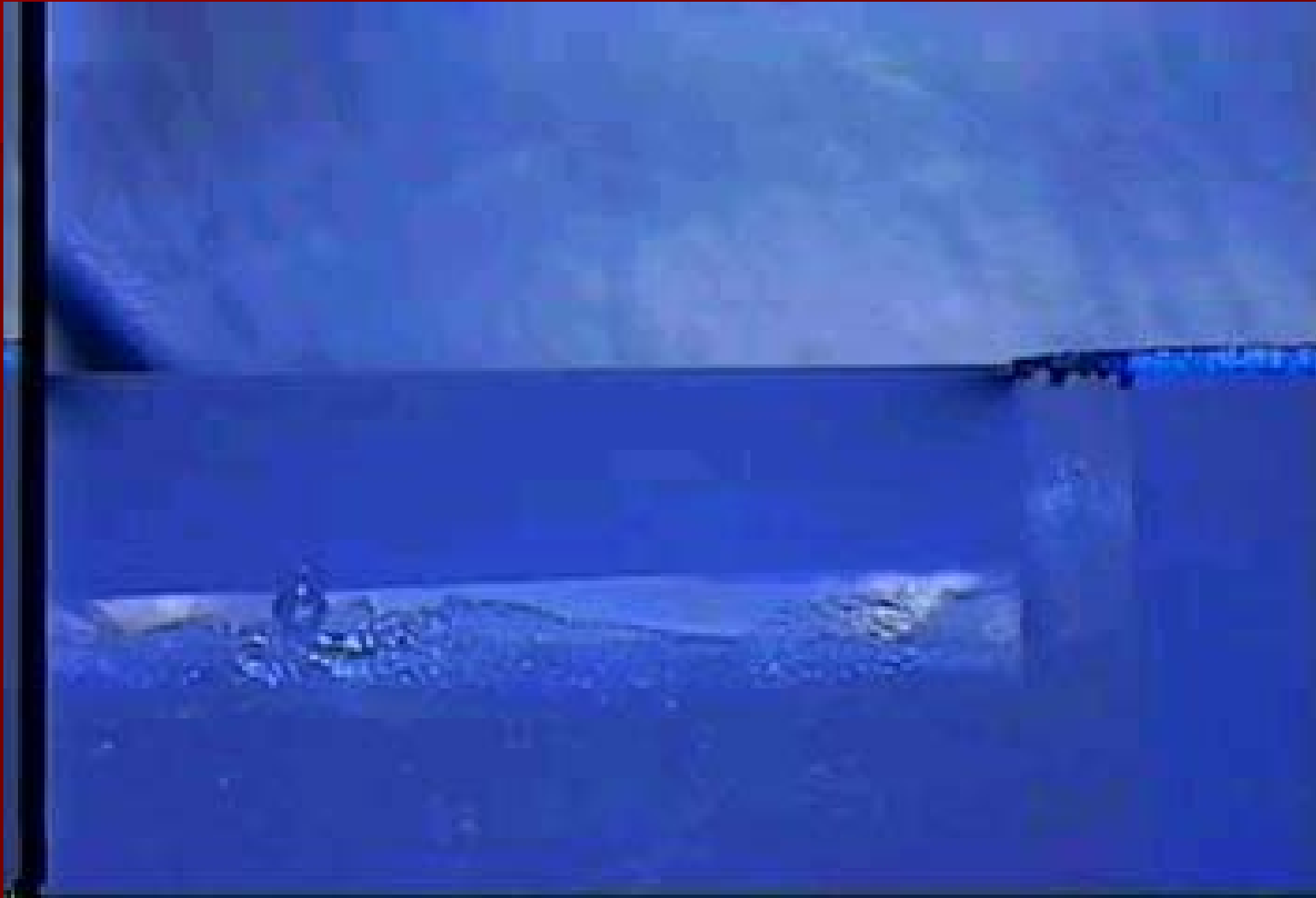


Diatome CryoTrim 45° Tool for Precise trimming of the Trapezoid

# Position diamond knife for serial sectioning



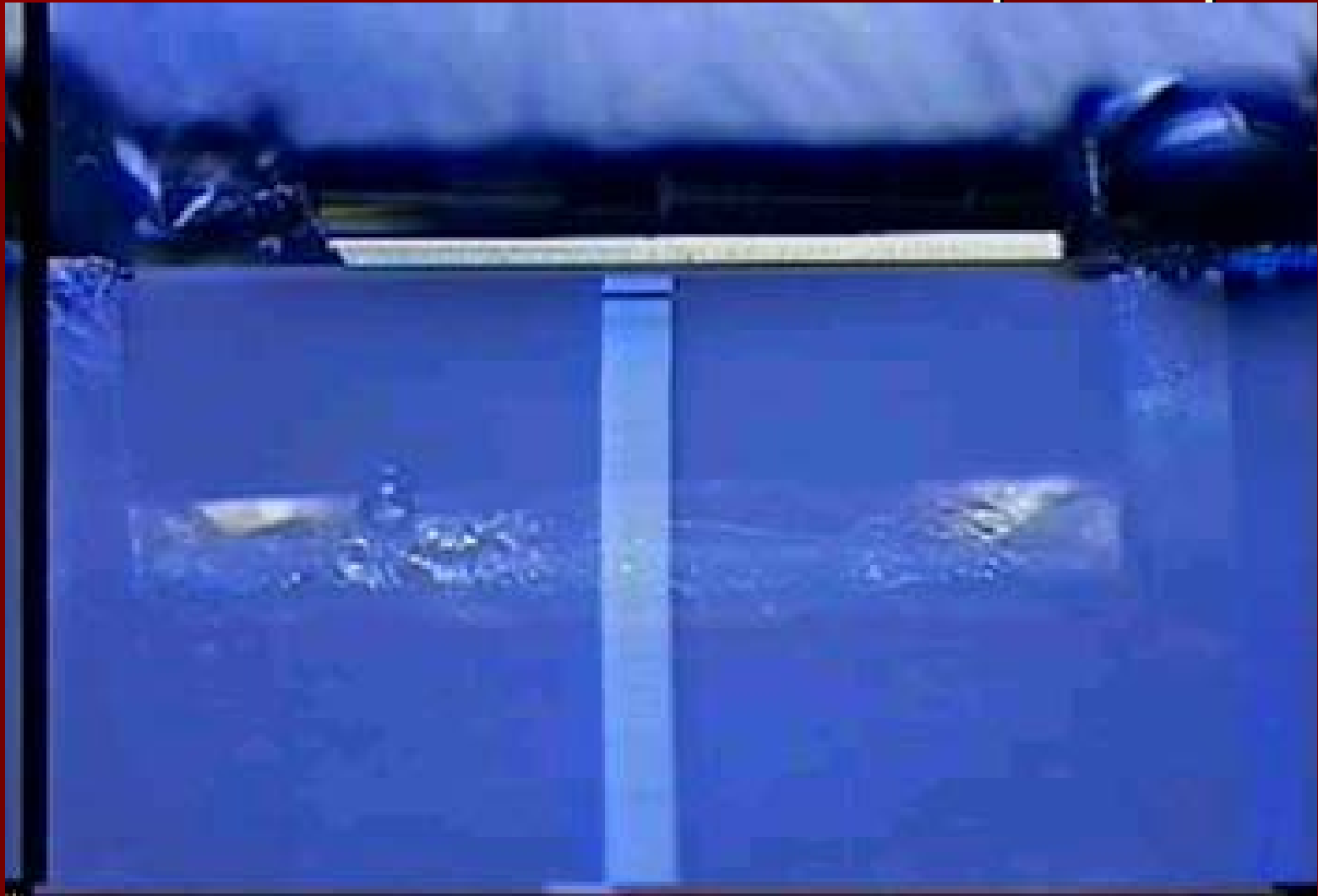
# Ribbon coming off of Knife



Sectioning rate: 1 mm / second; 40 – 50 nm thickness

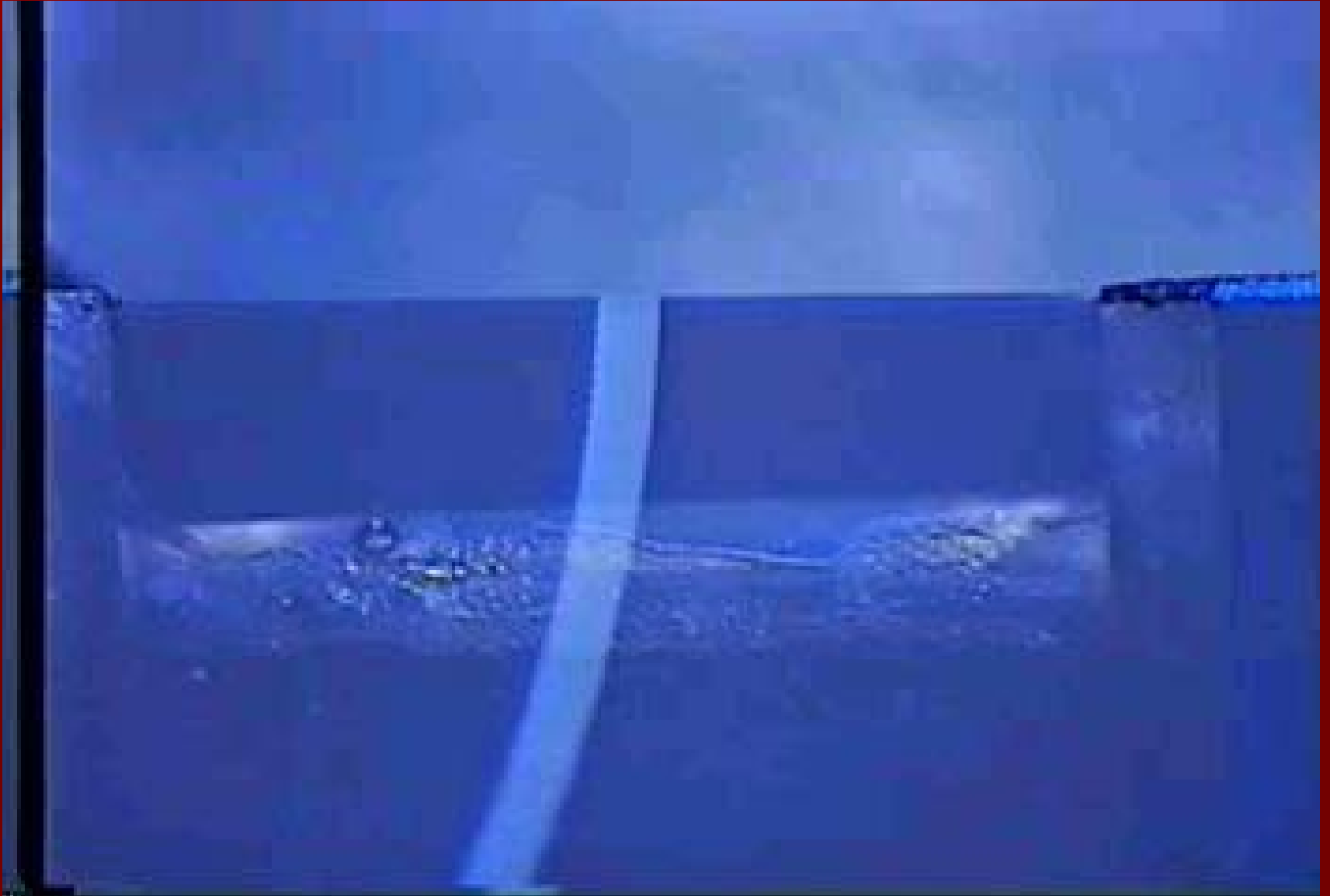


# 420 section series ribbon/pick up



The Perfect Eyelash Set available from EMS, Cat. #70616-10

# Broken Series pick up



# Grid staining, Washing, WICKING DRY!



# Loading grid to Gimbol





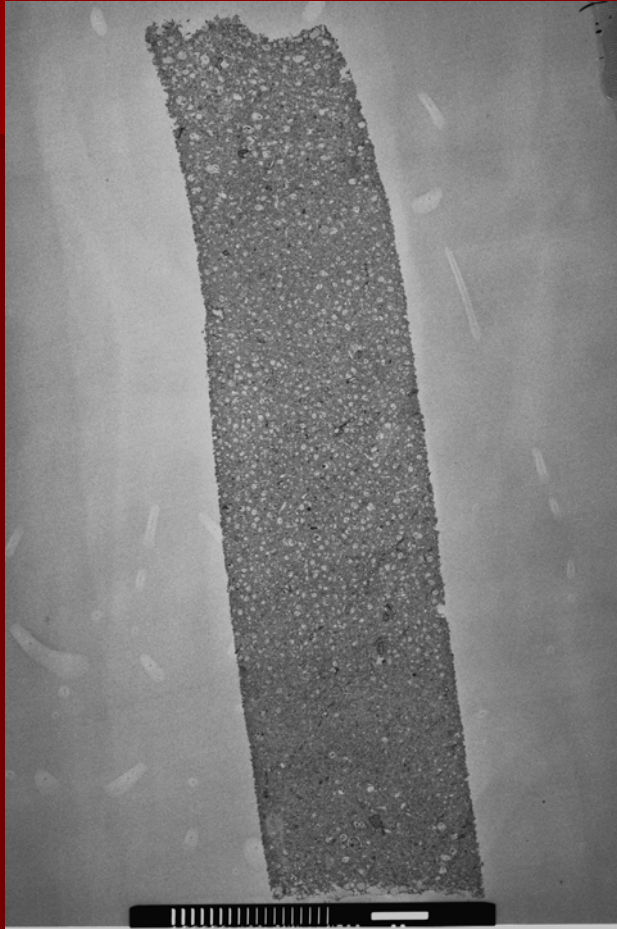
# Loading to a Rotational Holder



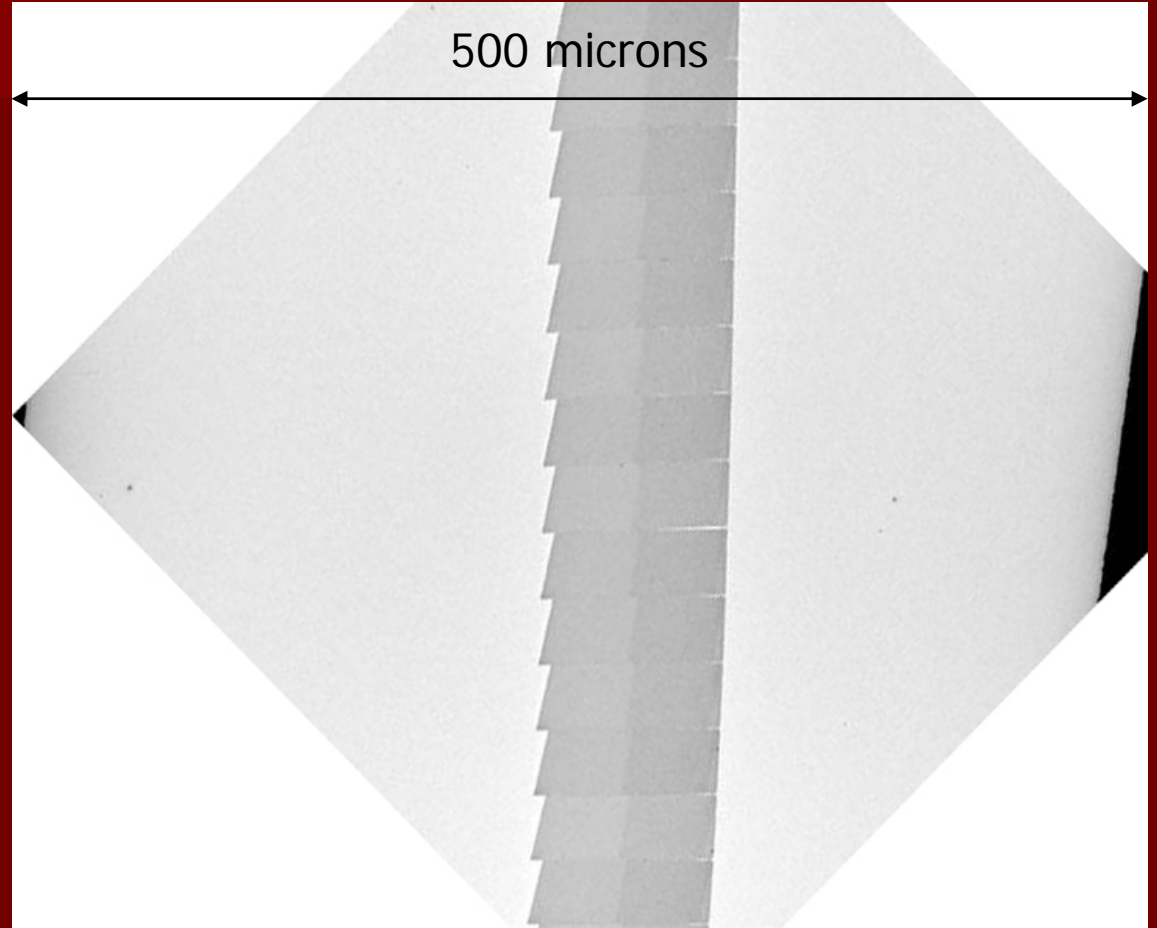
# Digital photography JEOL 1230 with 4K Gatan



# Fold-free, clean, uniform sections



Test Thins



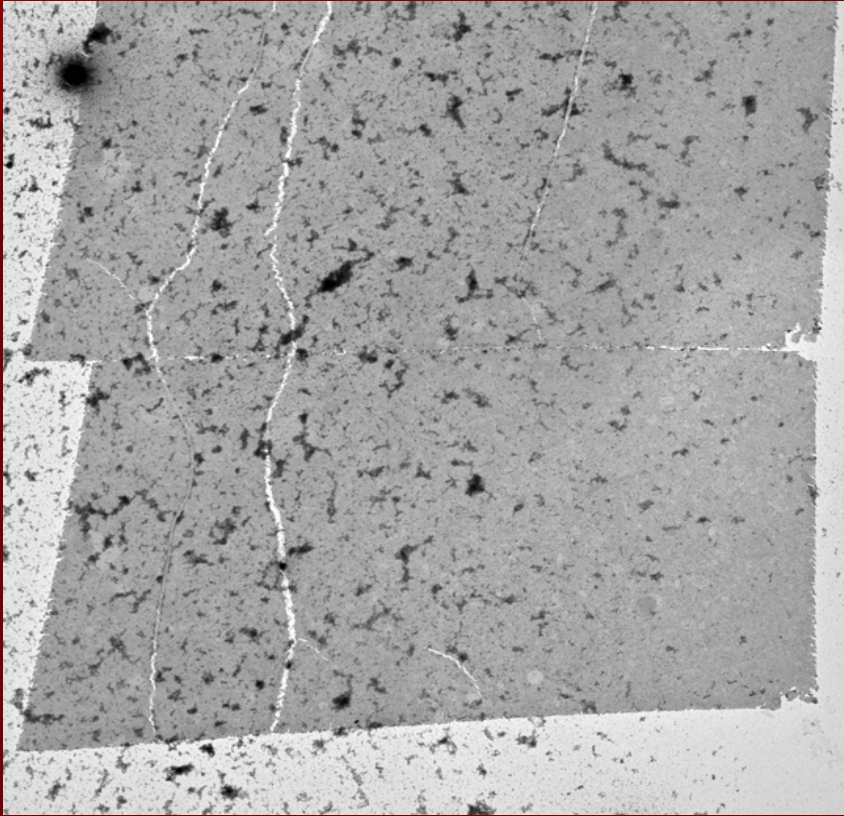
Serial Sections

# PITFALLS!

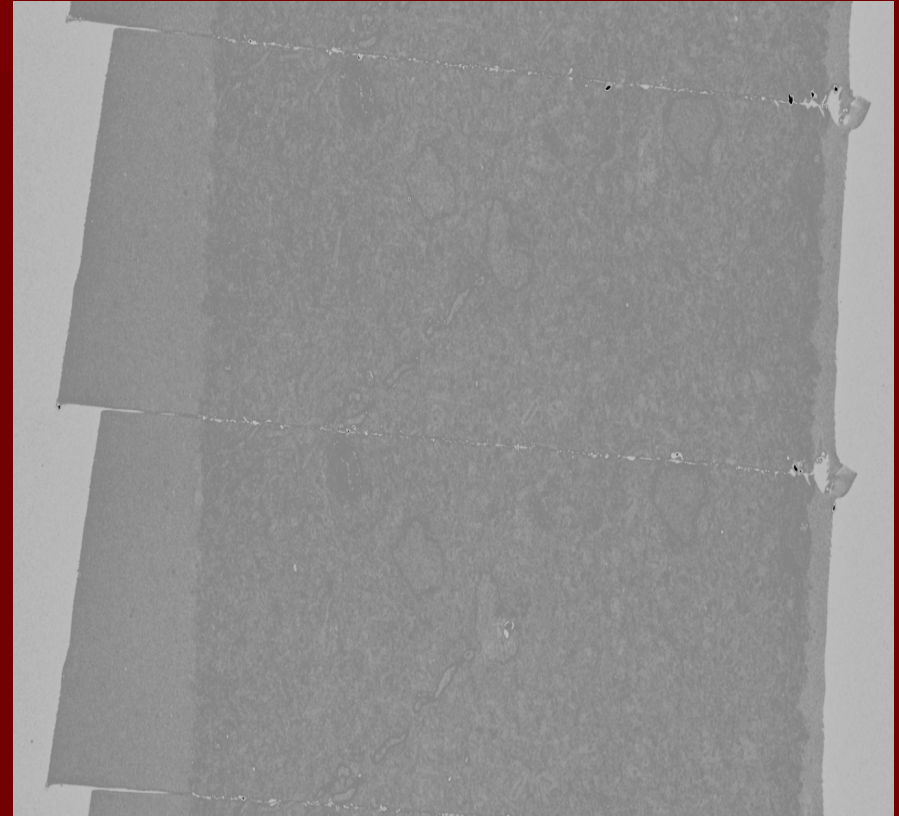
And solutions... 😊



# Dirty old stains, or improperly dried

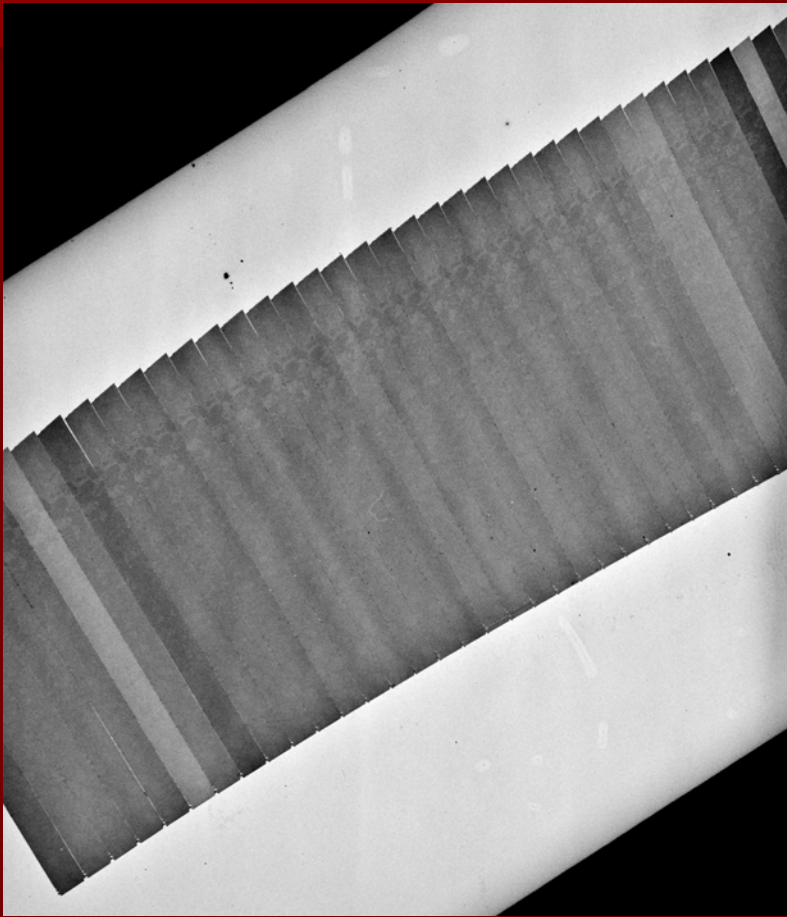


Lead Citrate Stain Precipitate



Make fresh stain often  
to have clean sections

# Uneven section thickness



- Eliminate Airdrafts!
- Eliminate Heat variation
  - Body temperature
    - Get out of the room!
- Ensure parallel top and bottom trimming
- Perfect Epon curing
- Make knife angle parallel to block face
- Etc...

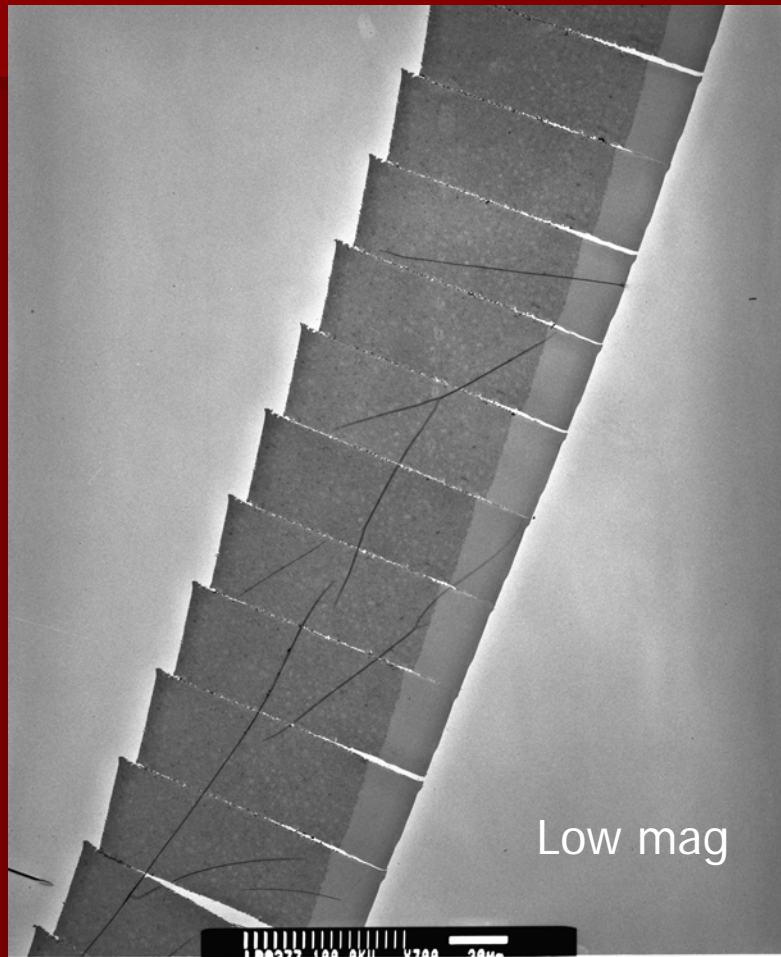
# Graphite plexiglas enclosure fixes most uneven sections problems



Here enclosing the Leica NEW UC6 ultramicrotome



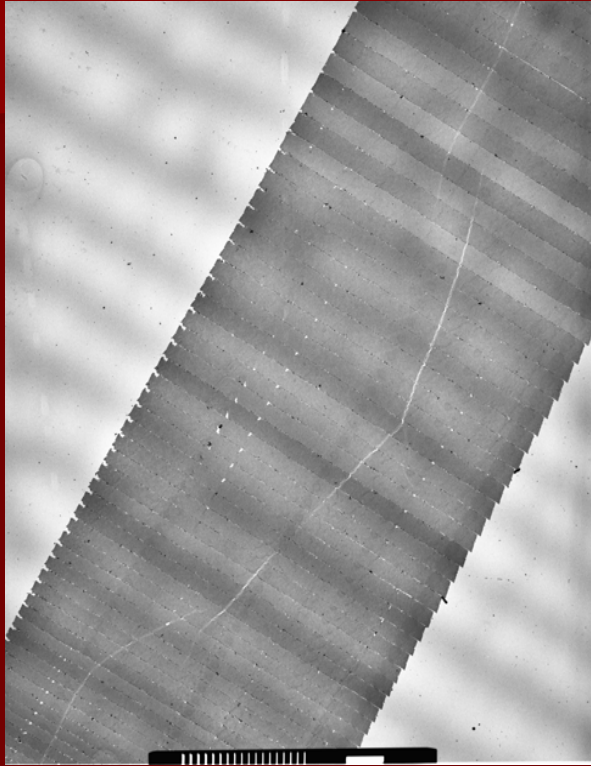
# Folds, Ugh! – >1 day Old Pioloform Slot grids



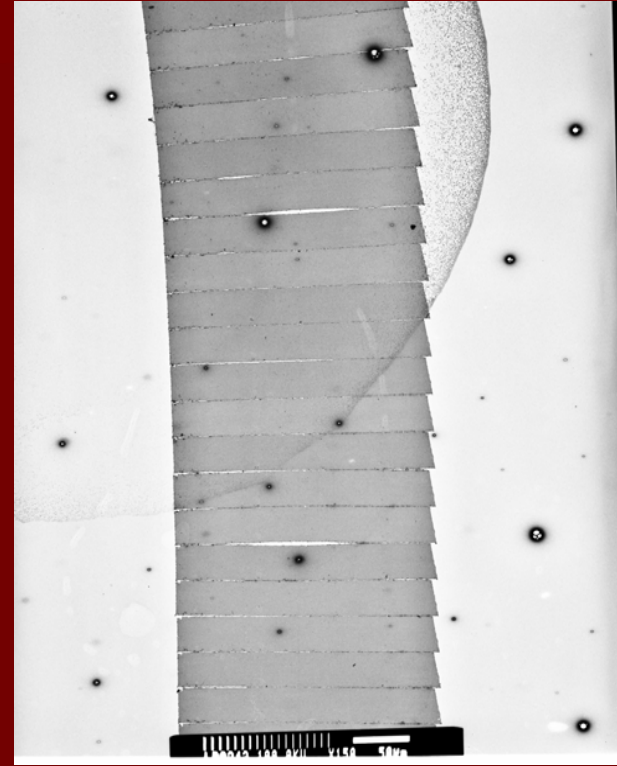
Folds caused by section drying down on a saggy Pioloform film.



## Other Grid Coating Problems

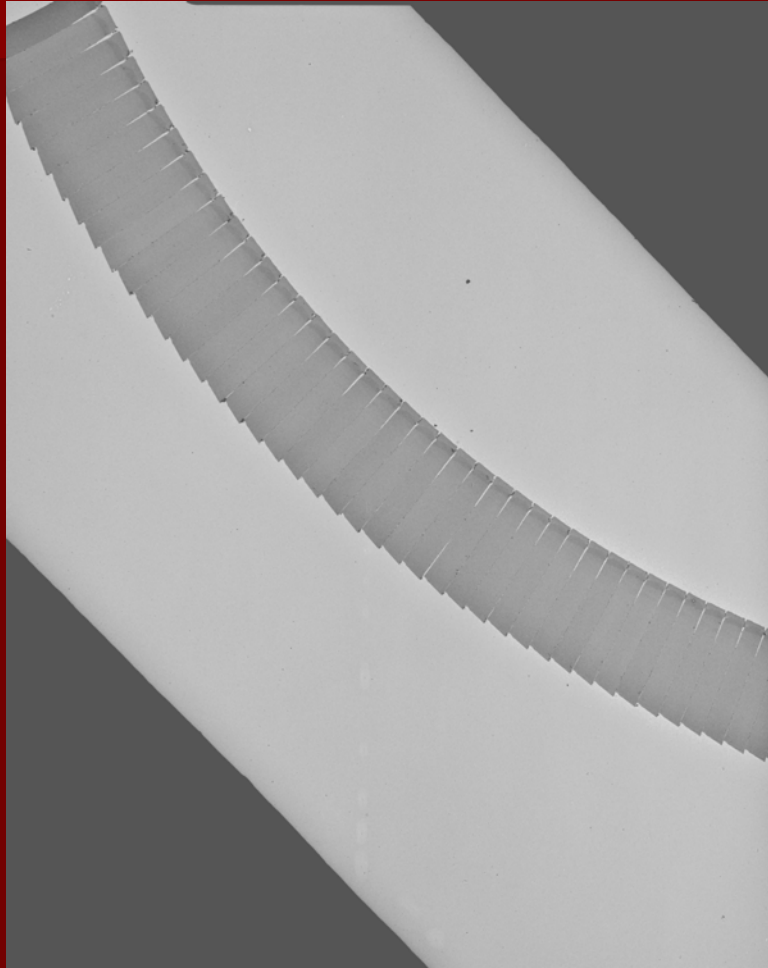


Uneven Pioloform  
Too Thick coats unevenly  
(Gold not Sliver  
Interference Color)



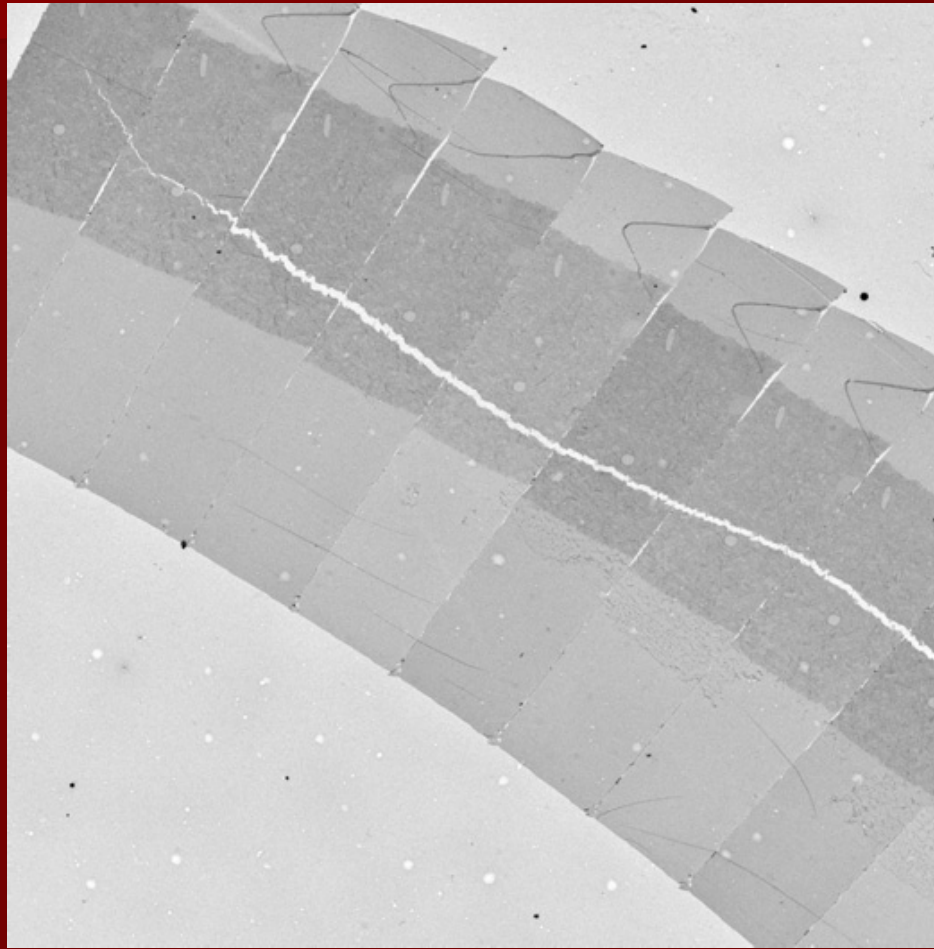
Pioloform  
Holes Caused by  
moisture

# Curved Ribbon – hard to center on the narrow slot grids



Non-parallel top and bottom

# Tissue cracks



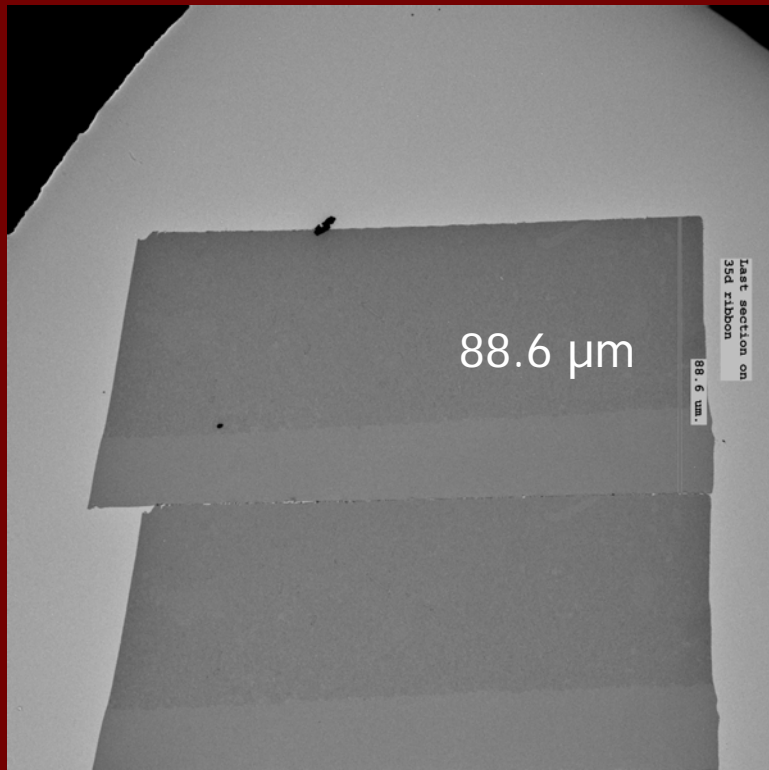
Poor Resin Infiltration

35° diamond knife for serial sectioning  
gives least compression

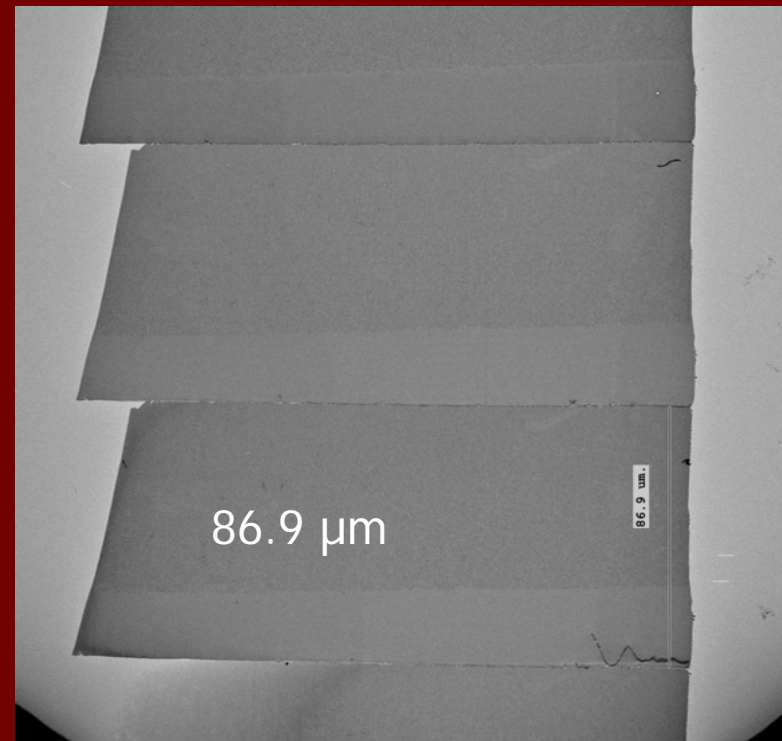




# Acute knife angles cause more compression



Section 200 cut with 35° Knife



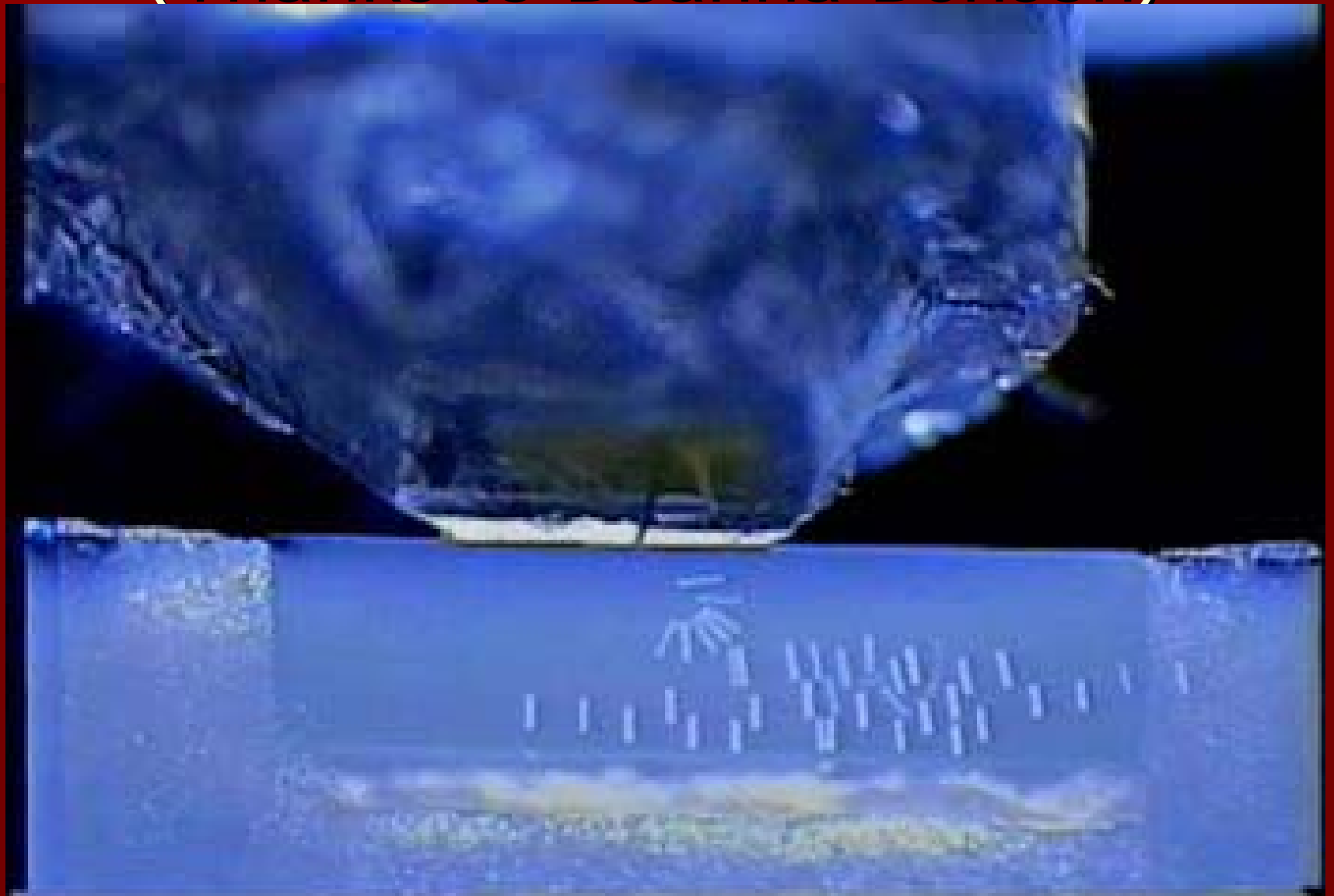
Section 201 in same series cut with 45° Knife  
Has 10% more compression

# How 'bout series and other Resins

- Lowicryl for post-embedding Immunogold labelling...
- Huge problems for 3D series – solved last week!

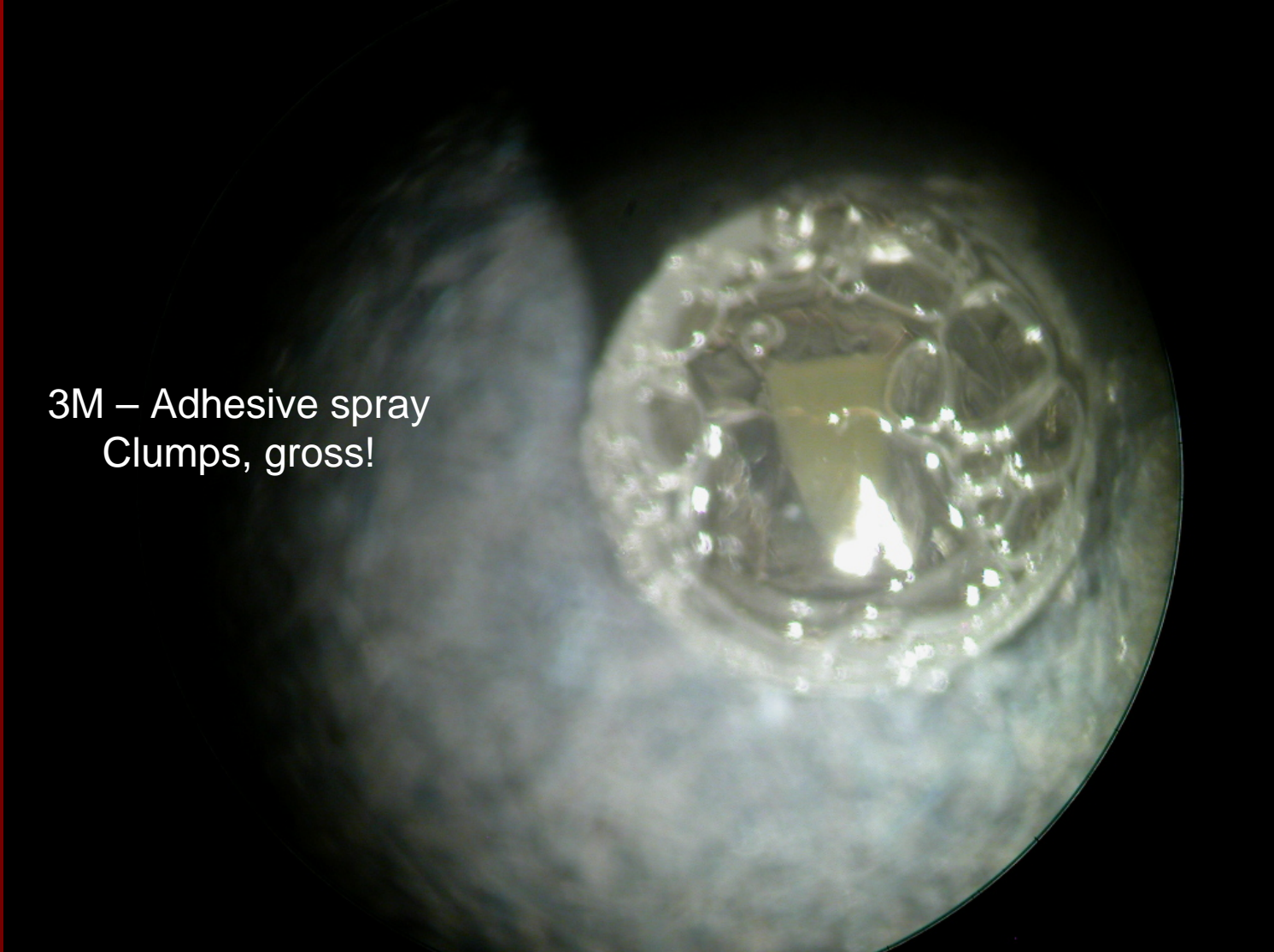
# Lowicryl

(Thanks to Deanna Benson)



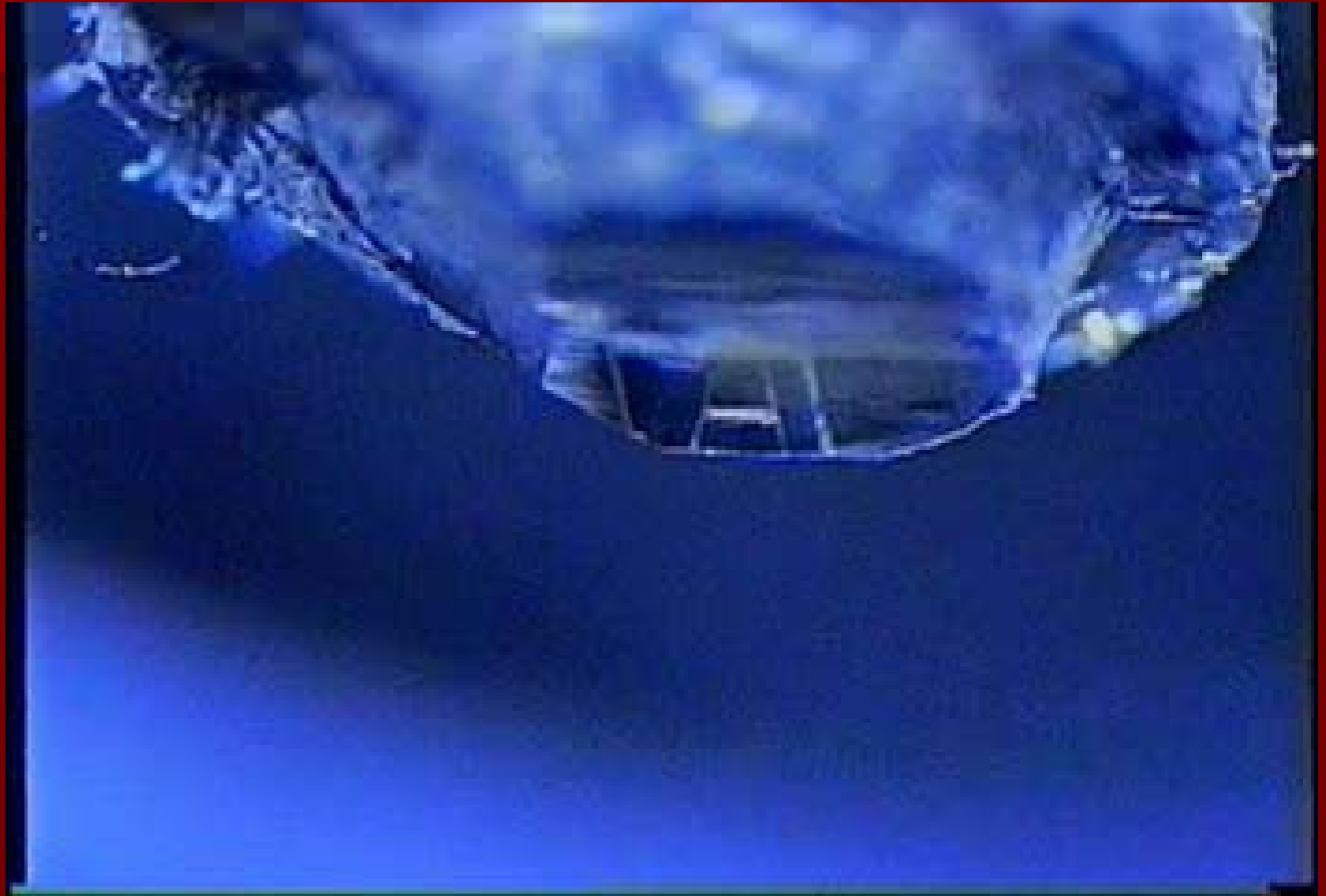
# Not Just Any Glue will do

3M – Adhesive spray  
Clumps, gross!

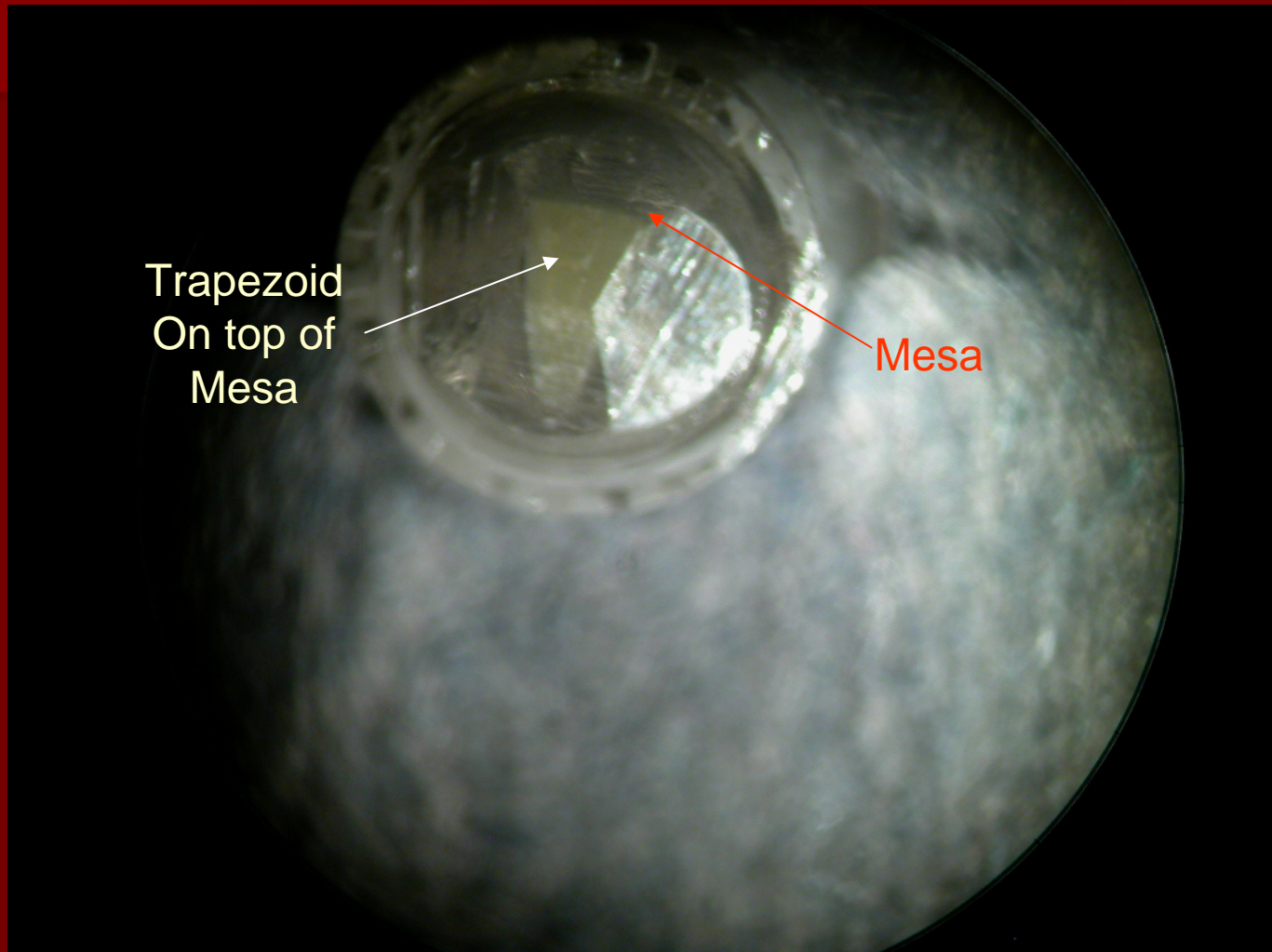


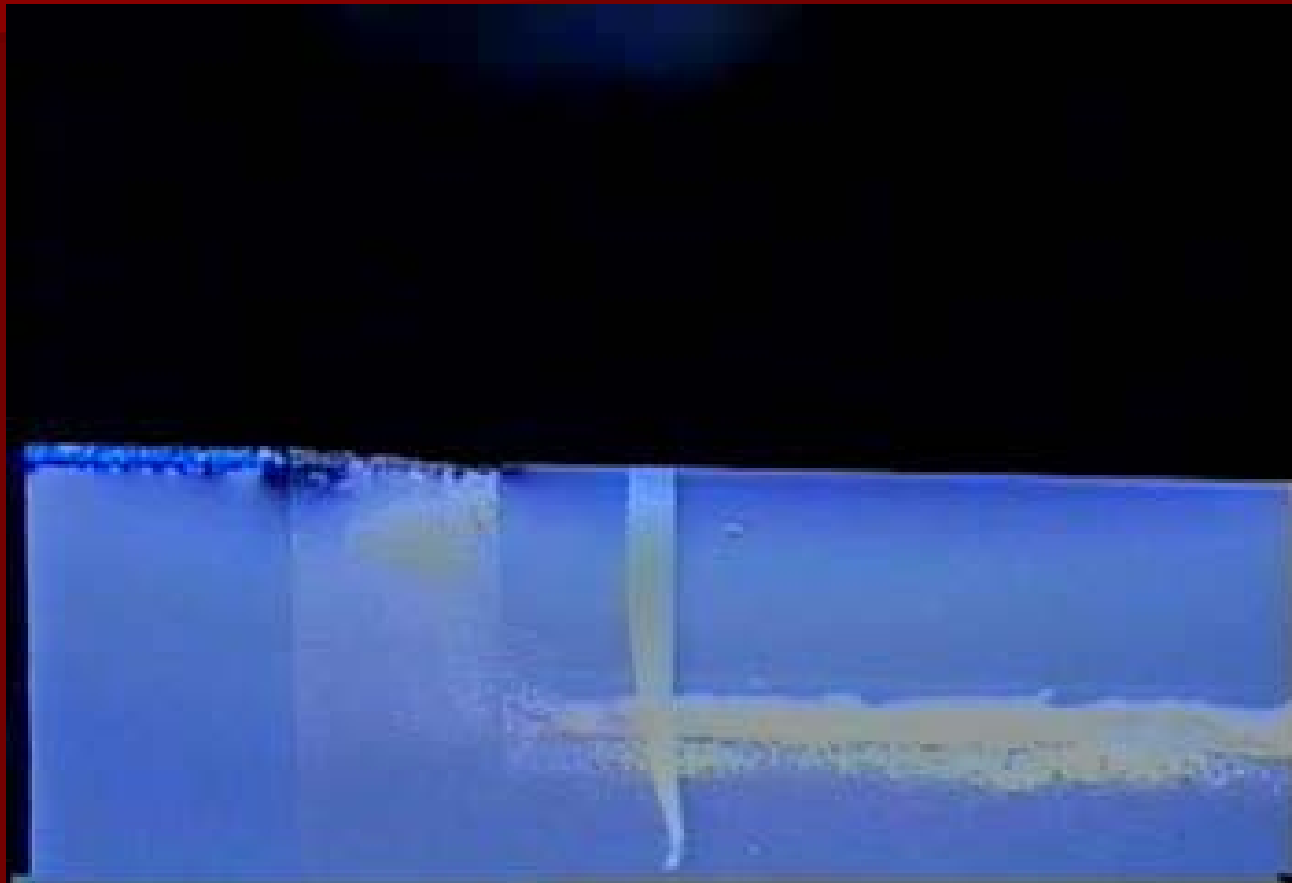


# Hair Salon Hair Spray



# Clean dry hair spray



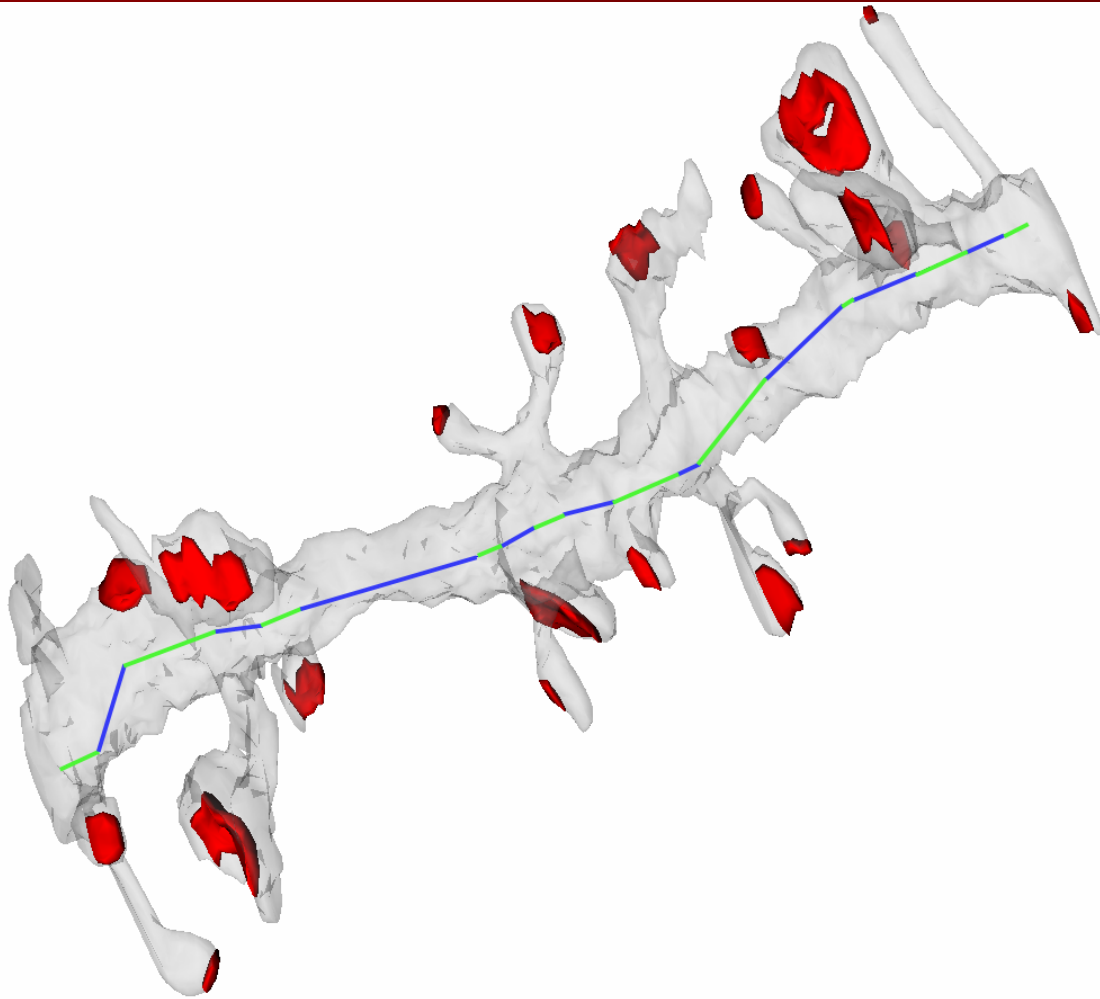


# Posters with new data - Harris Lab

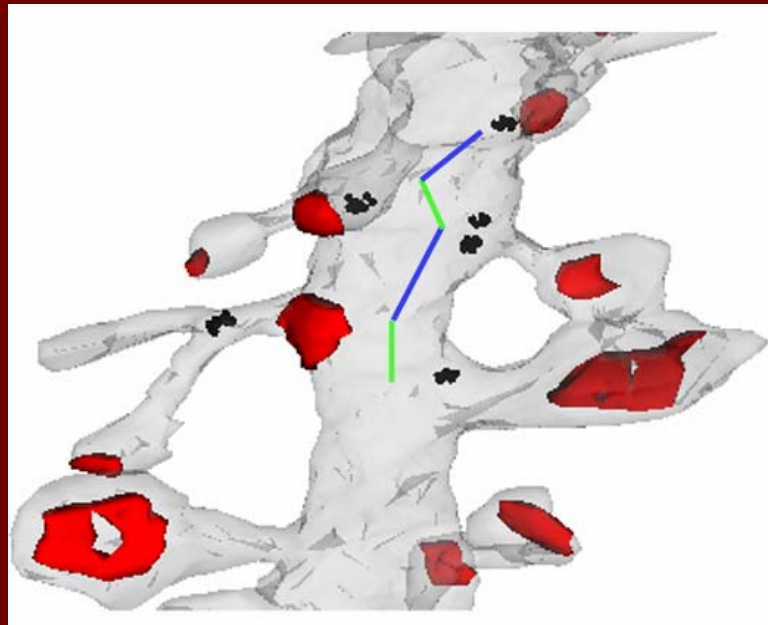
- 1:00 Sat. #43.18 (Mishra et al)
  - Dense core vesicles as substrate for synaptogenesis in hippocampus
- 8:00 Mon. #385.1 (Witcher et al)
  - Plasticity of Perisynaptic astroglial process at mature hippocampal synapses
- 1:00 Mon. #501.2 (Bourne and Harris)
  - Old title: Spinules and LTP at mature CA3->CA1 synapses
  - New Title: Structural Plasticity of Mature CA3-> CA1 synapses during LTP
- 1:00 Mon. #570.5 (Shi et al.)
  - 3d Editor – new software development



# Linear Next Neighbor Analysis: A Powerful Measure of Spine Density



Detect Polyribosome clustering  
Most PR closer together in LTP



So how do we do this?

# RECONSTRUCT

John Fiala

Download from: [synapses.bu.edu](http://synapses.bu.edu)



# Reconstruction through Serial EM

- Example from LO 114c
  - Start section 55
  - Go up – stop at 67 to illustrate PR at a synapse
  - Go up to second synapse
  - Do 3D sects 55-86
  - D01 then d01c04a3D and d01c04b3d
  - Change to spheres and add the PR in this protrusion  
d01rh04a



# L0114 test sections

- Demonstrate image quality correction
  - Domain – adjust contrast
  - Domain attributes – see new values for brightness and contrast (B/C)
  - Apply these B/C values to all Sections
- Demonstrate alignment
  - Images are digital from a 4K X 4K Gatan camera on the JEOL 1230 EM
  - Hence, pretty well-aligned from the scope
  - RECONSTRUCT – bring into perfect alignment.
    - Sect 99 → 100 to start.
- Demonstrate calibration

# RECONSTRUCT – NQDBP sect 28

Demonstrate tracing and 3D – see also nice MSB.

The screenshot displays the NQDBP: 28 software interface. The main window shows a grayscale electron micrograph of a biological section. A large green rectangular box highlights a central region. Within this region, several structures are traced with colored outlines: a red outline for a small curved structure, a blue outline for a larger, more complex structure, and a yellow outline for another structure on the right. The 'Objects' list on the left side of the window contains the following data:

Object	Flat area
d16p08lin	0.00306876
d16p09	0.00304798
d16p10	0.0019053
d16p11	0.000266791
d16p12	0.0022502
d16p13	0.000628584
d16p14	0.000741633
d16p15	0.00133361
d16p16	0.000933336
d16p17	0.00106716
d16p18	0.0018858
d16p19	0.00304946
d16p21	0.000266667
d16p22	0.000343021
d16p23	0.000628075
d16p24	0.00296653
d16p25	0.000991088
d16p26	0.00925941
d16p27	0.000877216
d16p28	0.00141928
D17	33.393
d17mt	0.0324451
D18	12.4068
d18mt	0.0293666
D19	10.854
d19mt	0.0686528
mito1	0.0523671
mito10	0.0541994
mito11	0.0697444
mito12	0.0832829
mito13	0.0407206
mito14	0.0459894
mito15	0.0570743
mito16	0.0793585

The status bar at the bottom of the window indicates 'Section: 28', 'Slide 32 of 33', 'Slit', 'English (U.S.)', and a printer icon.